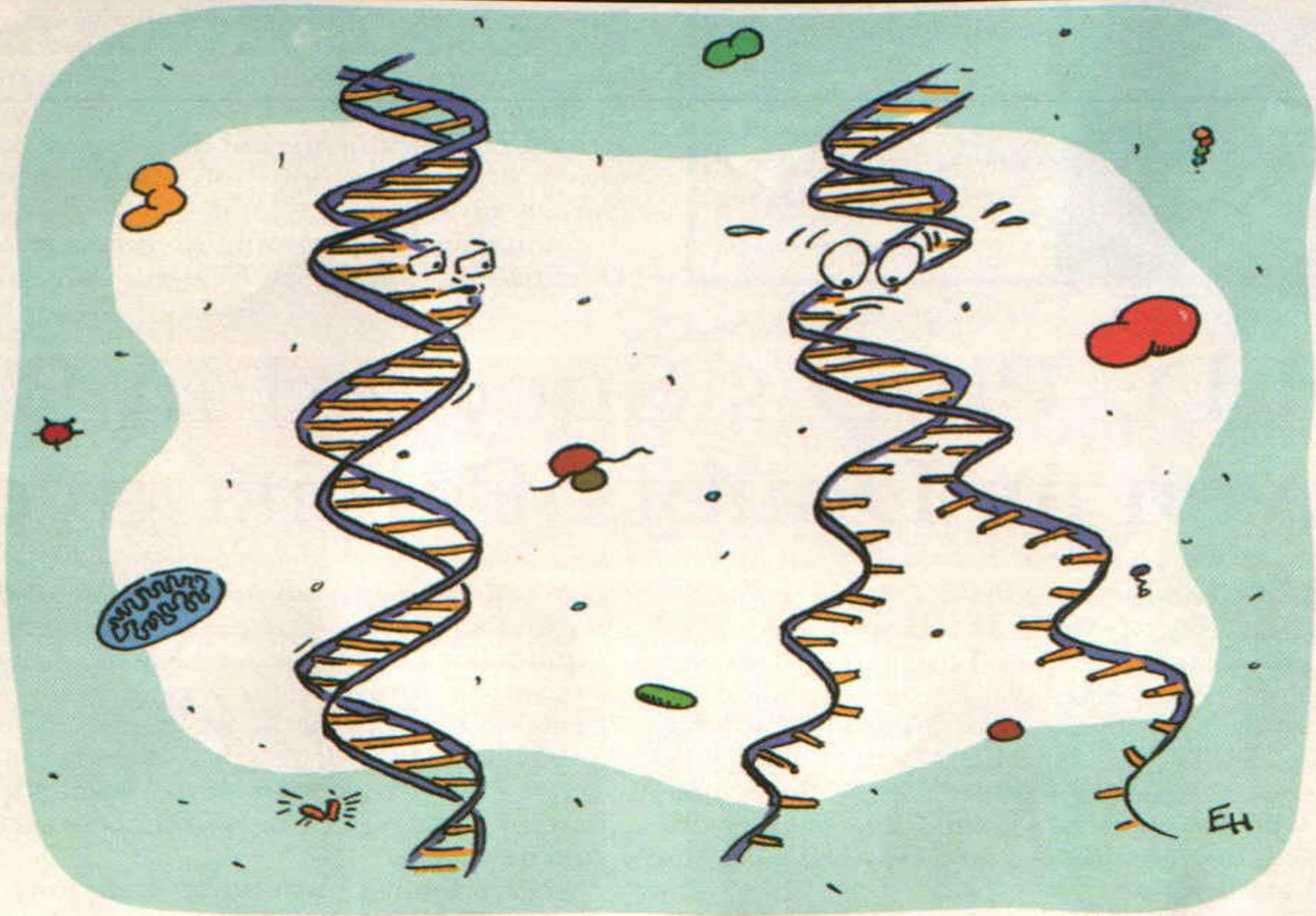


DNA Replication

Rasime Kalkan, PhD.

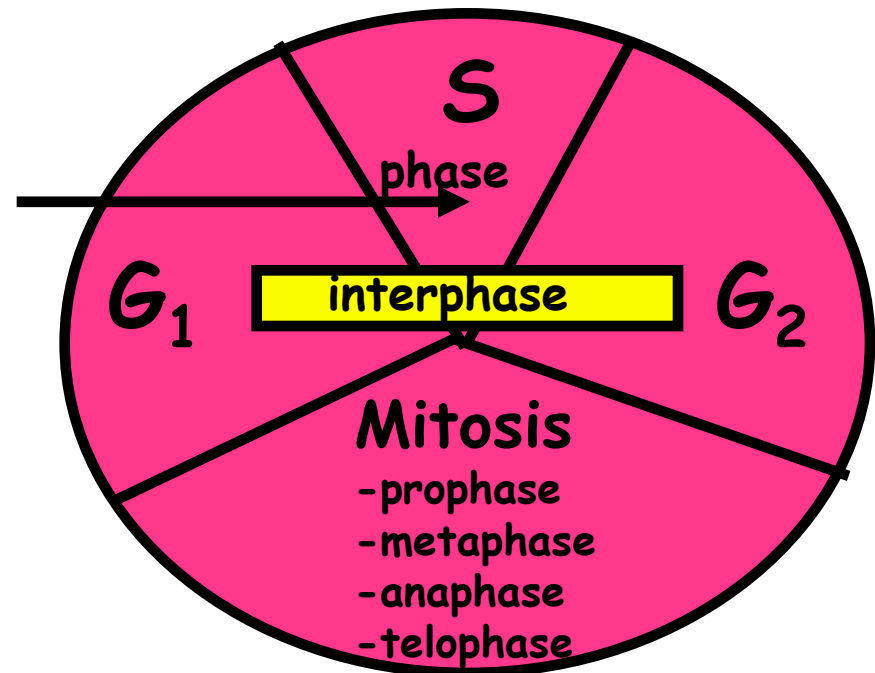


“Psst, Bob...you’re unzipped”

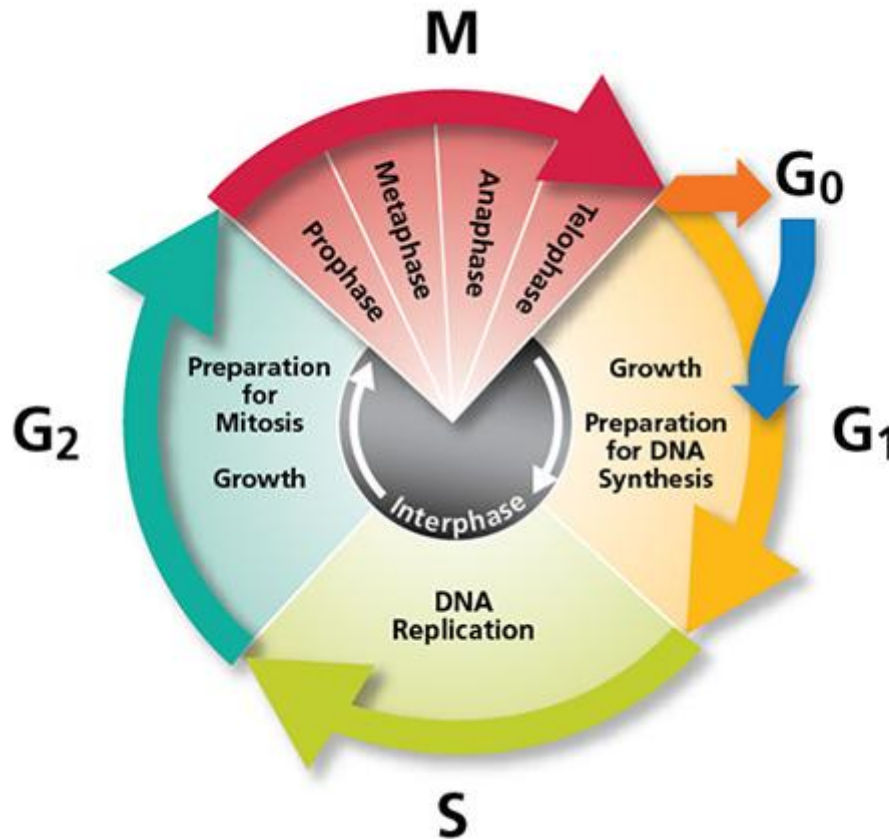
DNA Replication

- The process of copying one DNA molecule into two identical molecules is called **DNA replication**.
- DNA has to be copied before a cell divides
- DNA is copied during the S or synthesis phase of interphase
- New cells will need identical DNA strands

DNA replication takes place in the S phase.



The four standard phases of a eucaryotic cell
DNA replication occurring at S Phase (DNA synthesis phase)
G₁ and G₂, gap between S and M

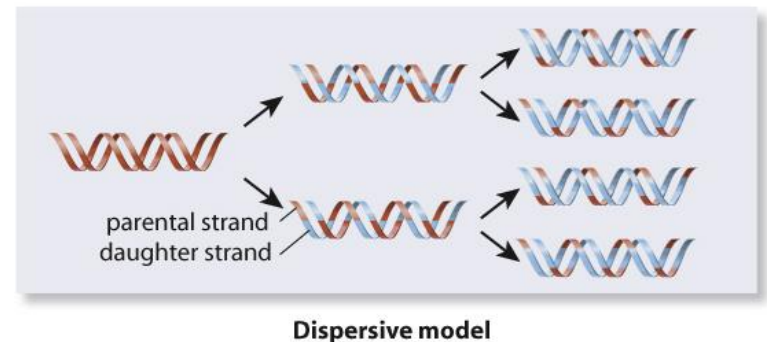
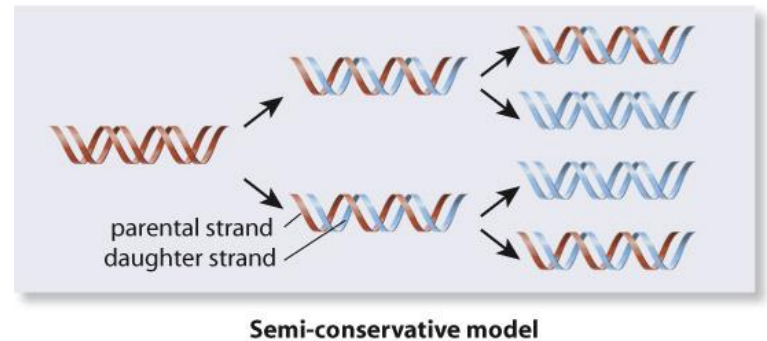
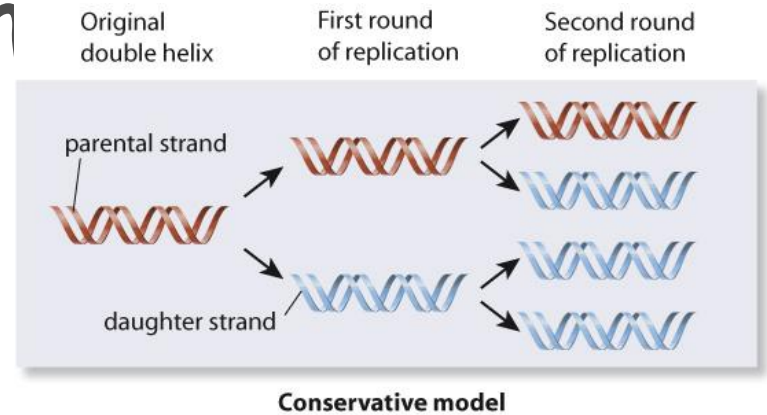


DNA Replication

In the mid-1950s, three competing models of DNA replication were proposed:

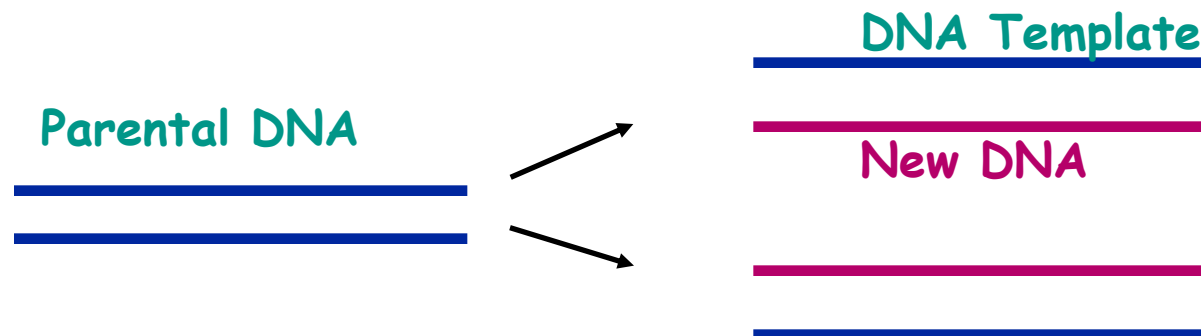
- conservative model
- semi-conservative model
- dispersive model

The conservative model results in one new molecule and conserves the old. The **semi-conservative replication** model results in two hybrid molecules of old and new strands. The dispersive model results in hybrid molecules with each strand being a mixture of old and new strands.



Semiconservative Model of Replication

- Idea presented by **Watson & Crick**
- The two strands of the parental molecule separate, and each acts as a **template** for a **new complementary strand**
- New DNA consists of 1 PARENTAL (original) and 1 NEW strand of DNA

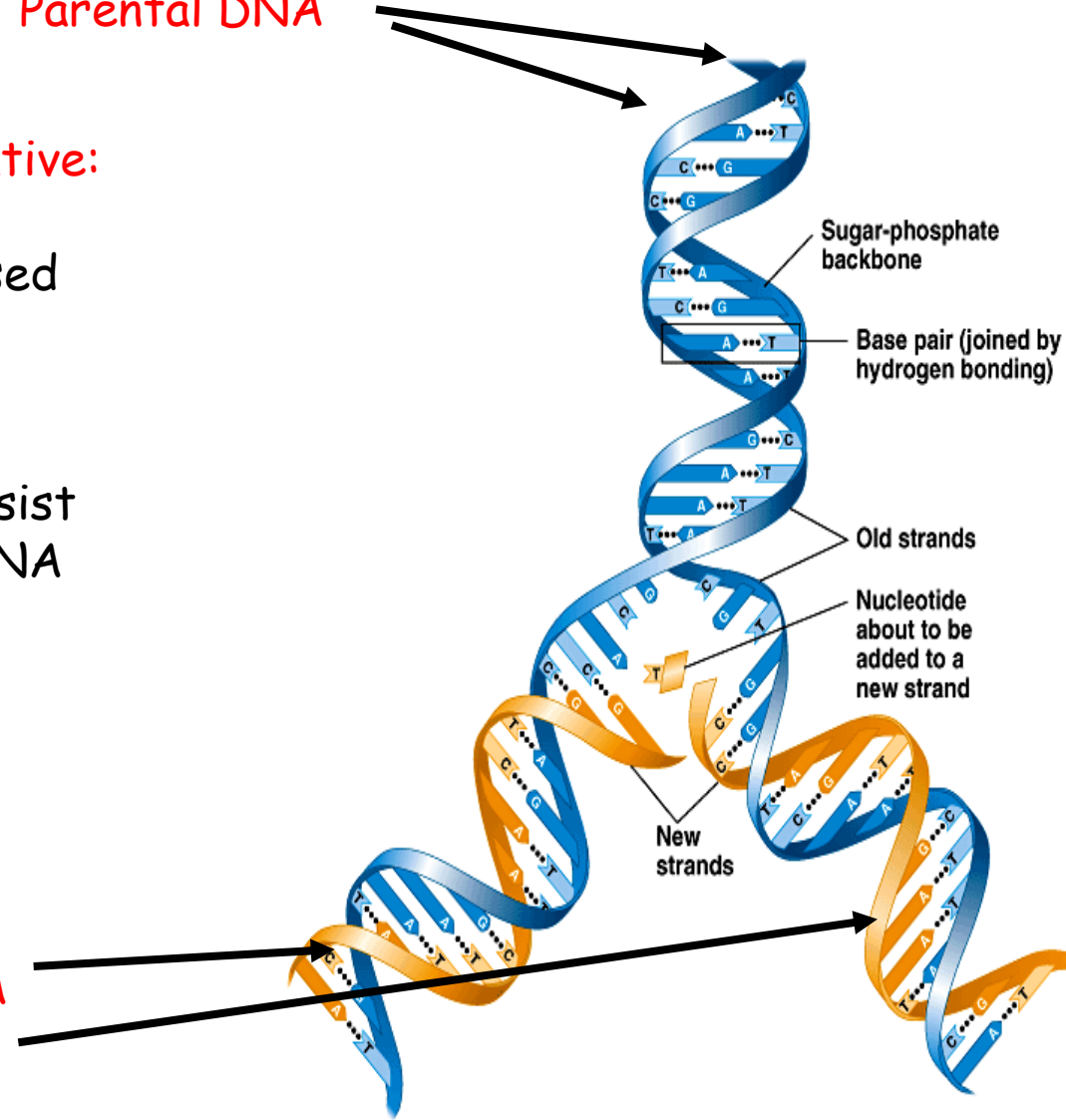


Parental DNA

DNA replication is semi conservative:

1. Parental DNA strands are used as a template for the newly replicated strands
2. Daughter DNA duplexes consist of one strand of parental DNA base paired to the newly replicated strand

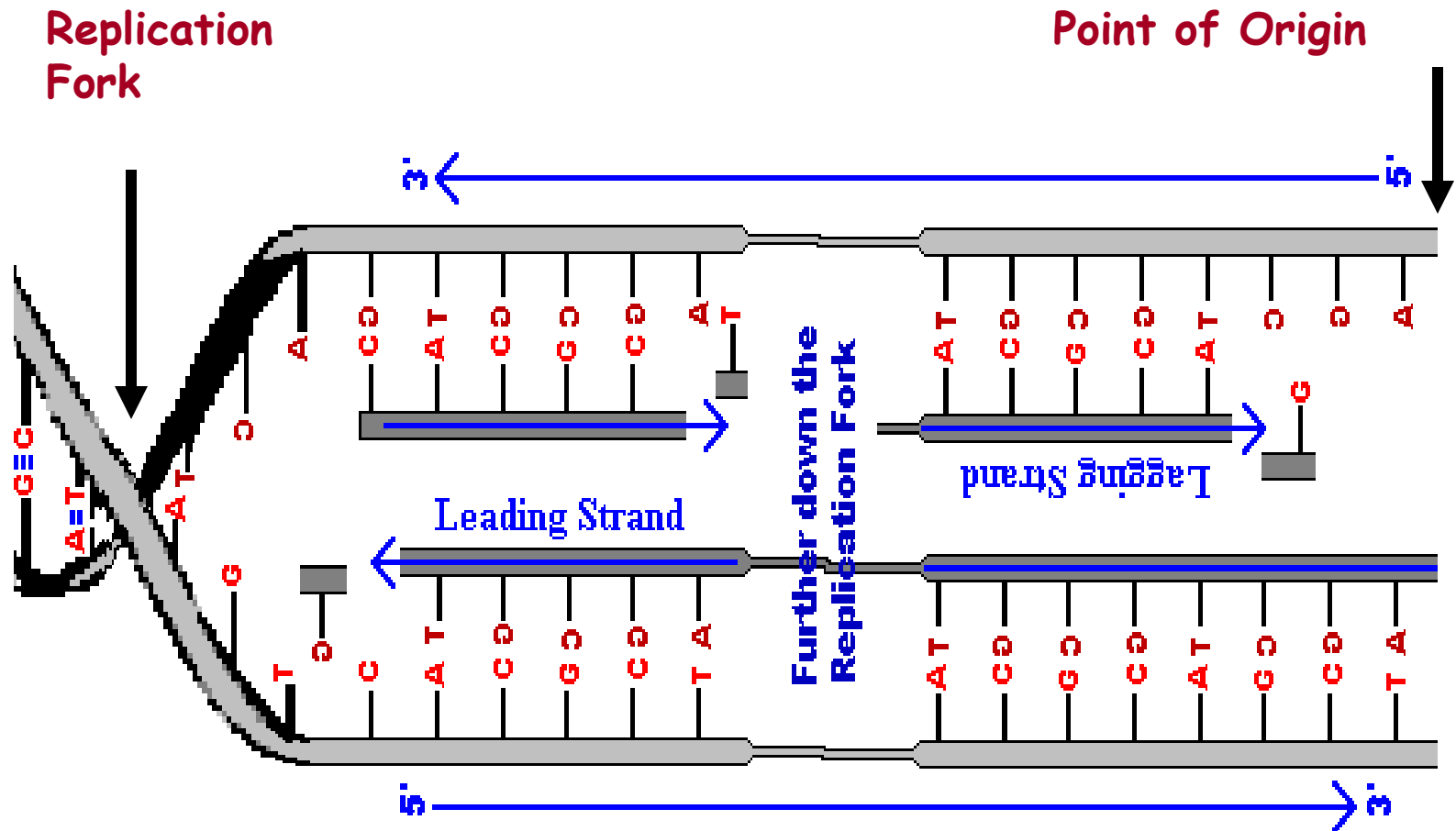
Newly replicated DNA strands



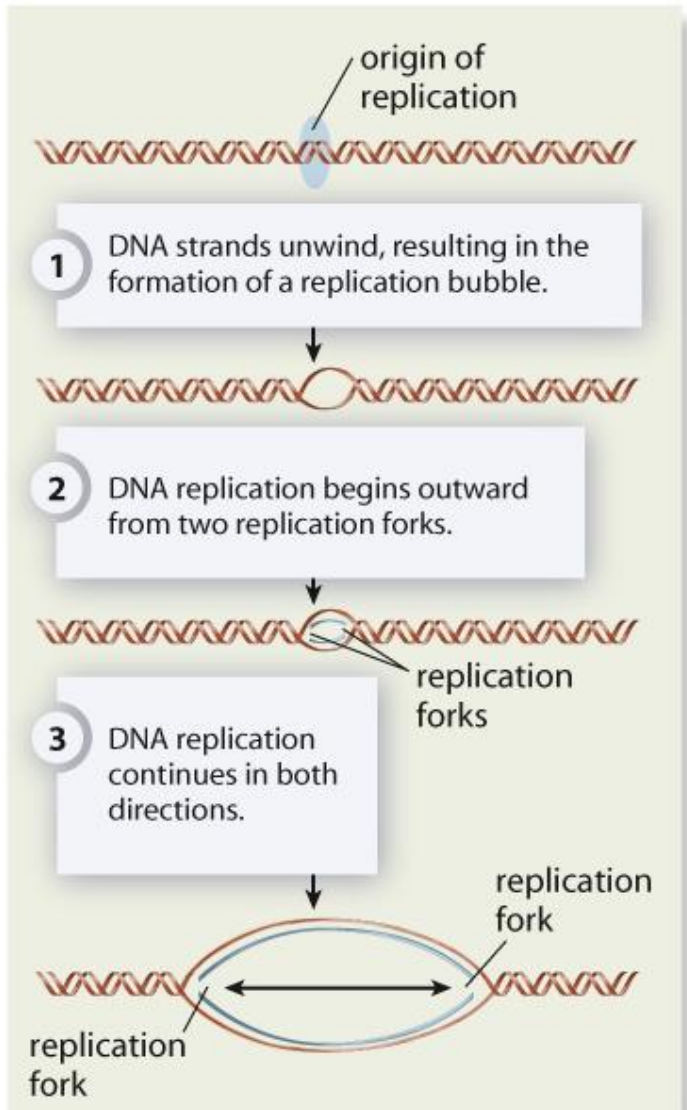
The mechanism of DNA replication

- Tightly controlled process,
 - occurs at specific times during the cell cycle.
 - Requires:
 - a set of proteins and enzymes,
 - and requires energy in the form of ATP.
- Two basic steps:
 - Initiation
 - Elongation.
 - Two basic components:
 - template
 - primer

Replication of Strands



DNA Replication



Semi-conservative replication has three phases: **initiation**, **elongation**, and **termination**.

DNA Replication

Topoisomerase - unwinds DNA

Helicase - enzyme that breaks H-bonds

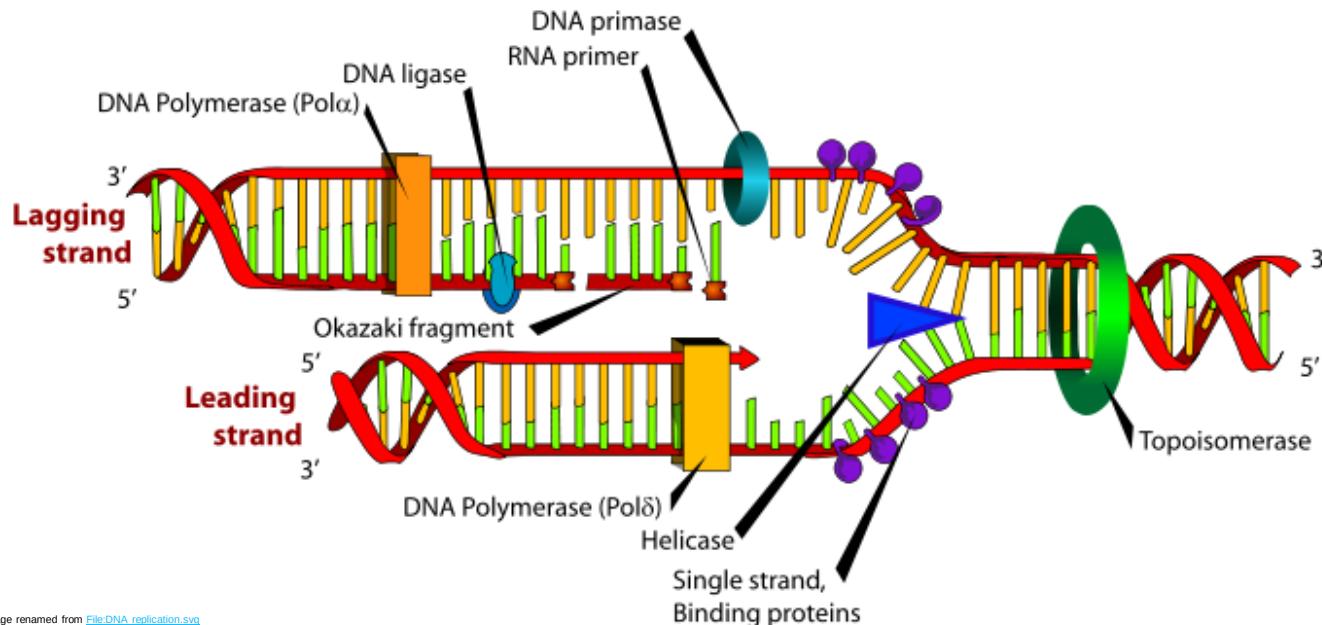
DNA Polymerase - enzyme that catalyzes connection of nucleotides to form complementary DNA strand in 5' to 3' direction (reads template in 3' to 5' direction)

Leading Strand - transcribed continuously in 5' to 3' direction

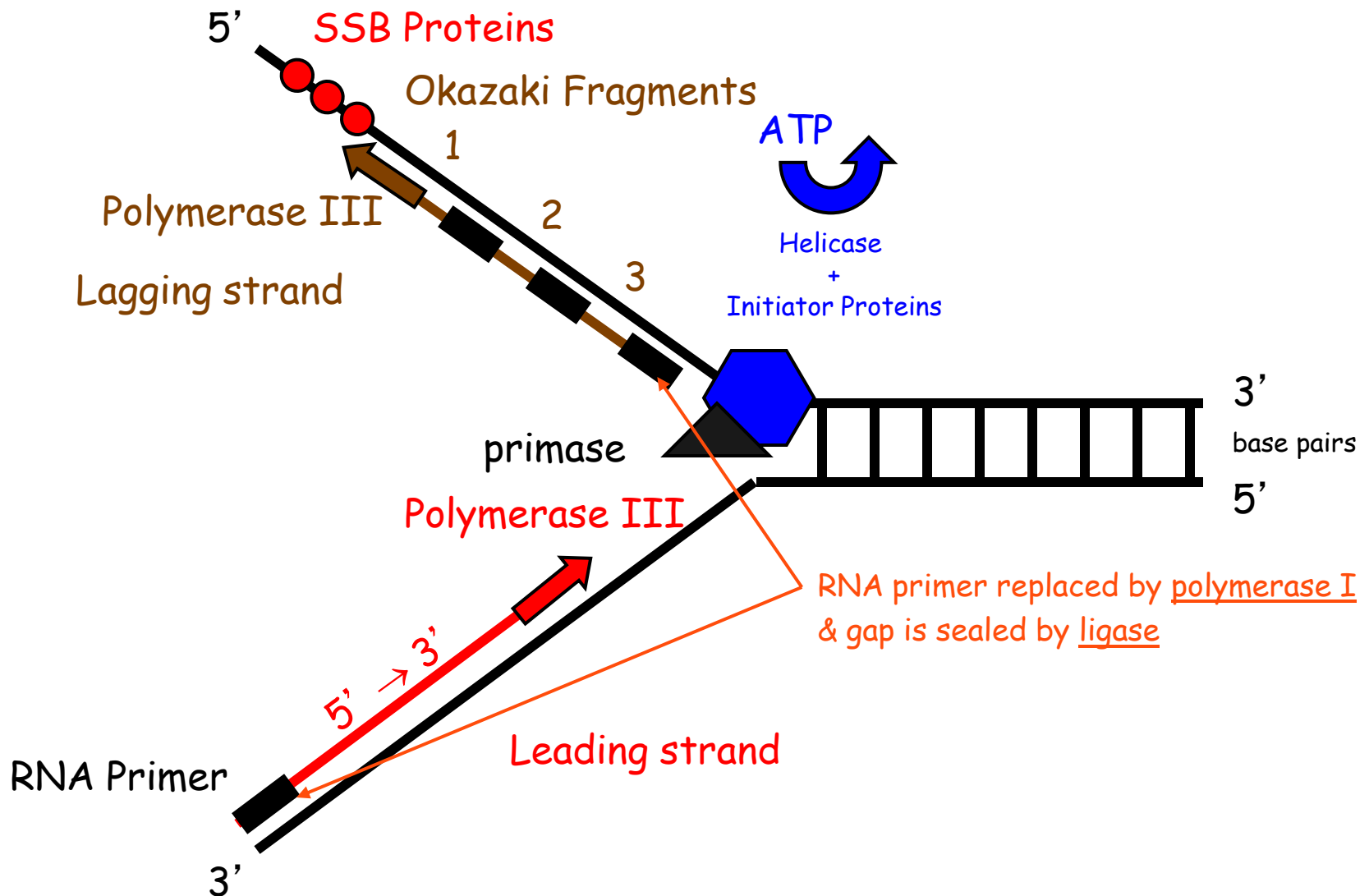
Lagging Strand - transcribed in segments in 5' to 3' direction (**Okazaki fragments**)

DNA Primase - enzyme that catalyzes formation of RNA starting segment (**RNA primer**)

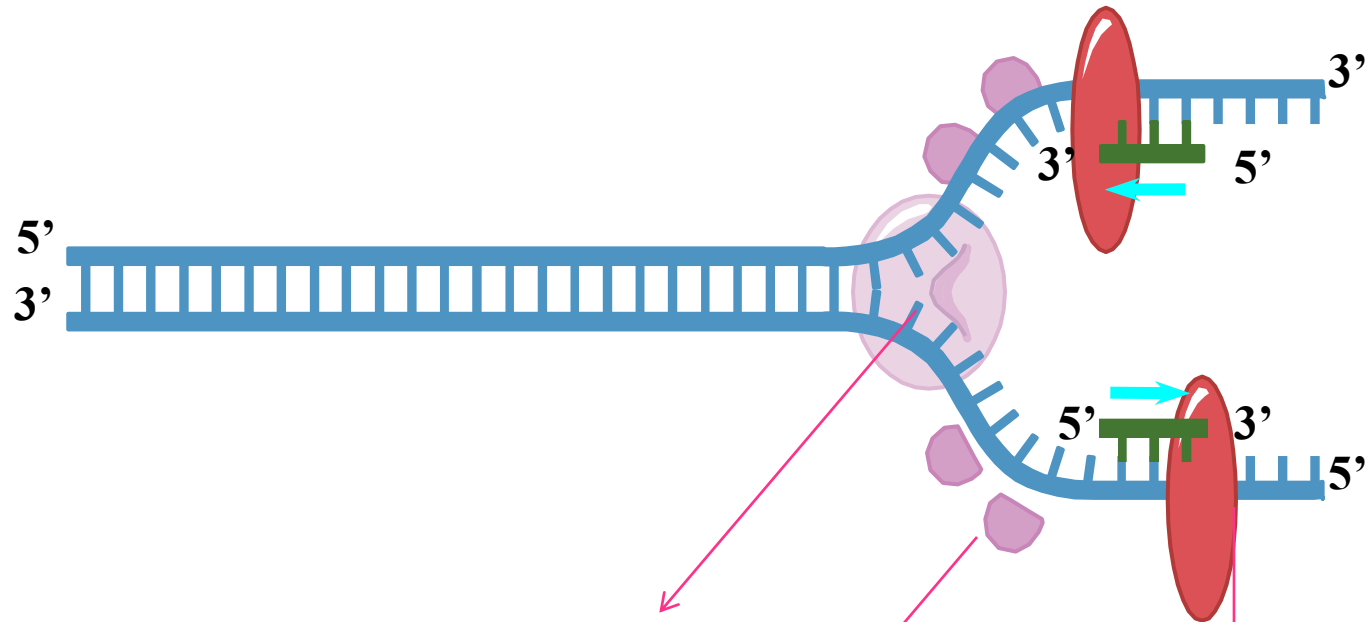
DNA Ligase - enzyme that catalyzes connection of two Okazaki fragments



Supercoiled DNA relaxed by gyrase & unwound by helicase + proteins:



Replication



Helicase protein binds to DNA sequences called origins and unwinds DNA strands.

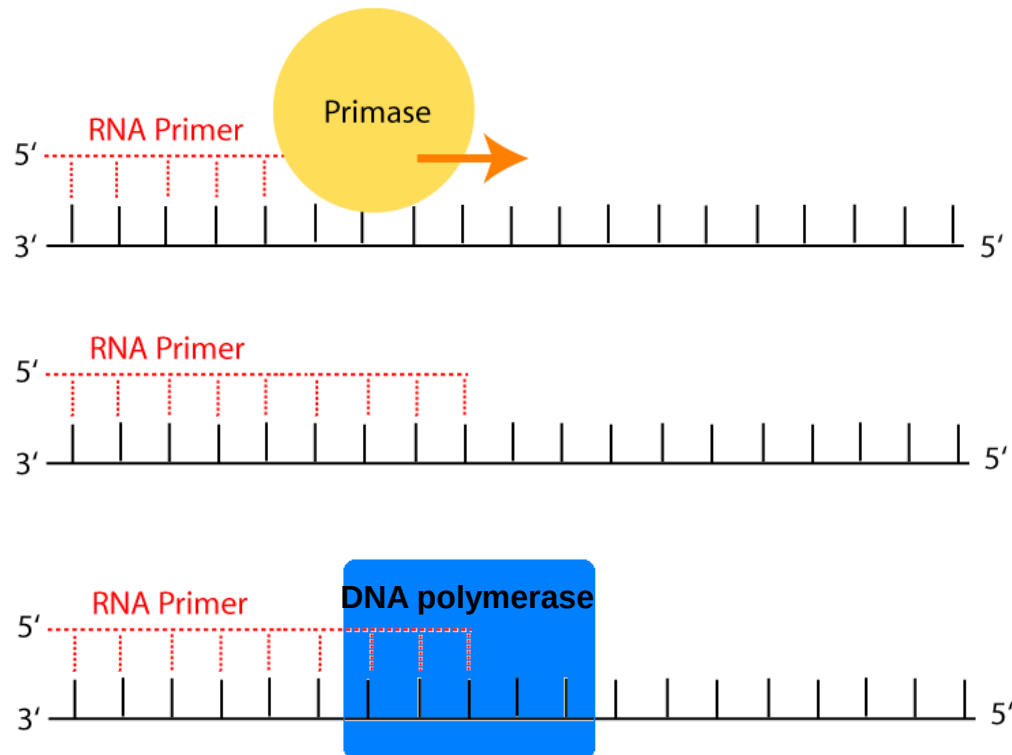
Binding proteins prevent single strands from rewinding.

Primase protein makes a short segment of RNA complementary to the DNA, a primer.

DNA Replication

Initiation

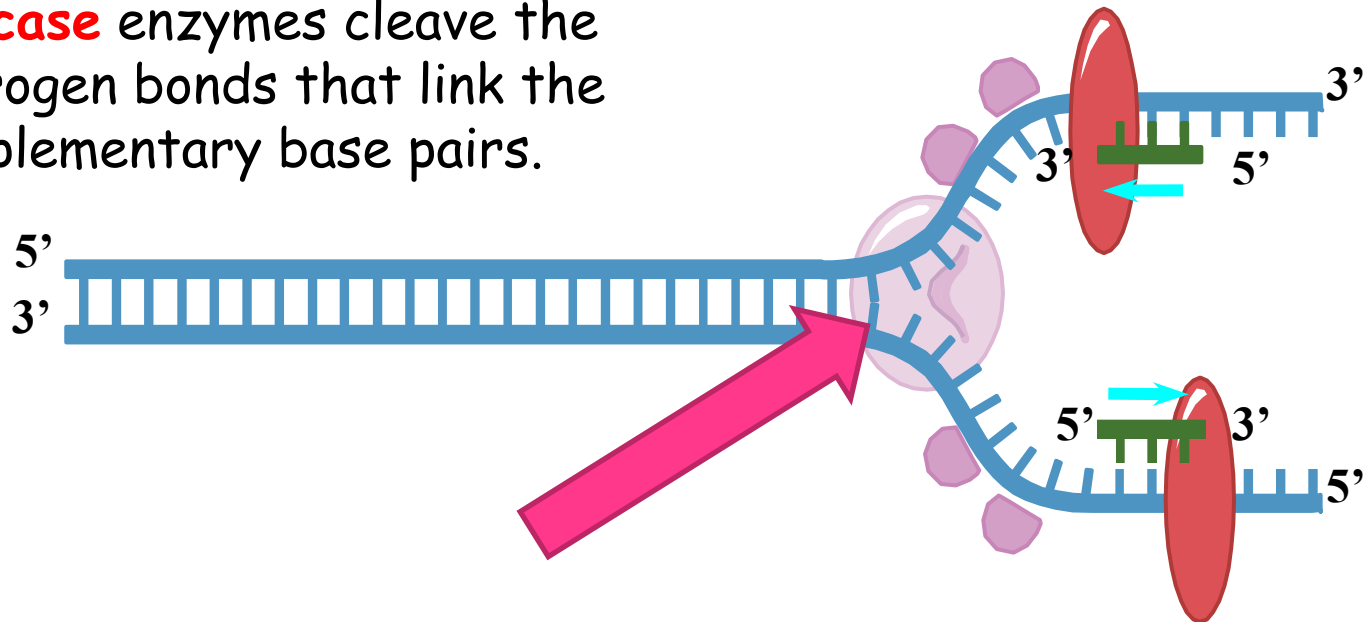
- **Primase** (a type of RNA polymerase) builds an **RNA primer** (5-10 ribonucleotides long)
- **DNA polymerase** attaches onto the 3' end of the **RNA primer**



Initiation of DNA replication

- Step 1 - opening the helix
 - Proteins bind to specific DNA sequences known as origins of replication
 - Bacteria have one
 - Eukaryotes have thousands
 - AT rich regions
 - - Why?
 - Helicases aid in the opening of the helix

- **Helicase** enzymes cleave the hydrogen bonds that link the complementary base pairs.



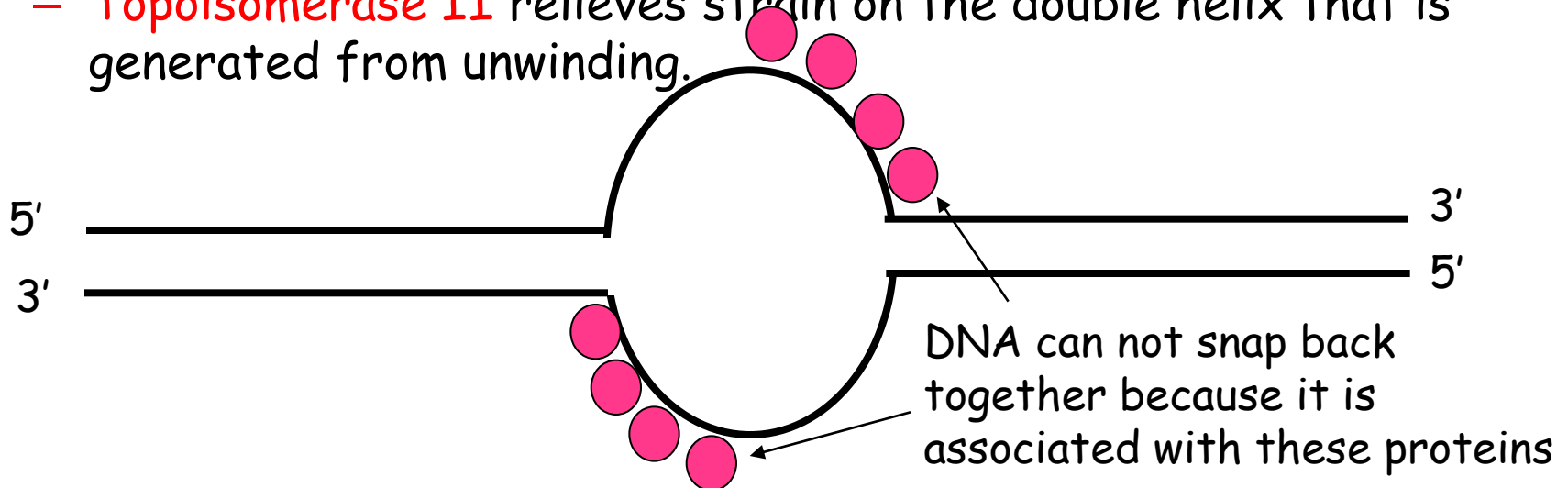
Initiation of DNA replication

- **Role of SSBP**

- Single stranded binding proteins
- After the helix has opened it is prevented from re-annealing by the action of these proteins
- These proteins stabilize single stranded DNA

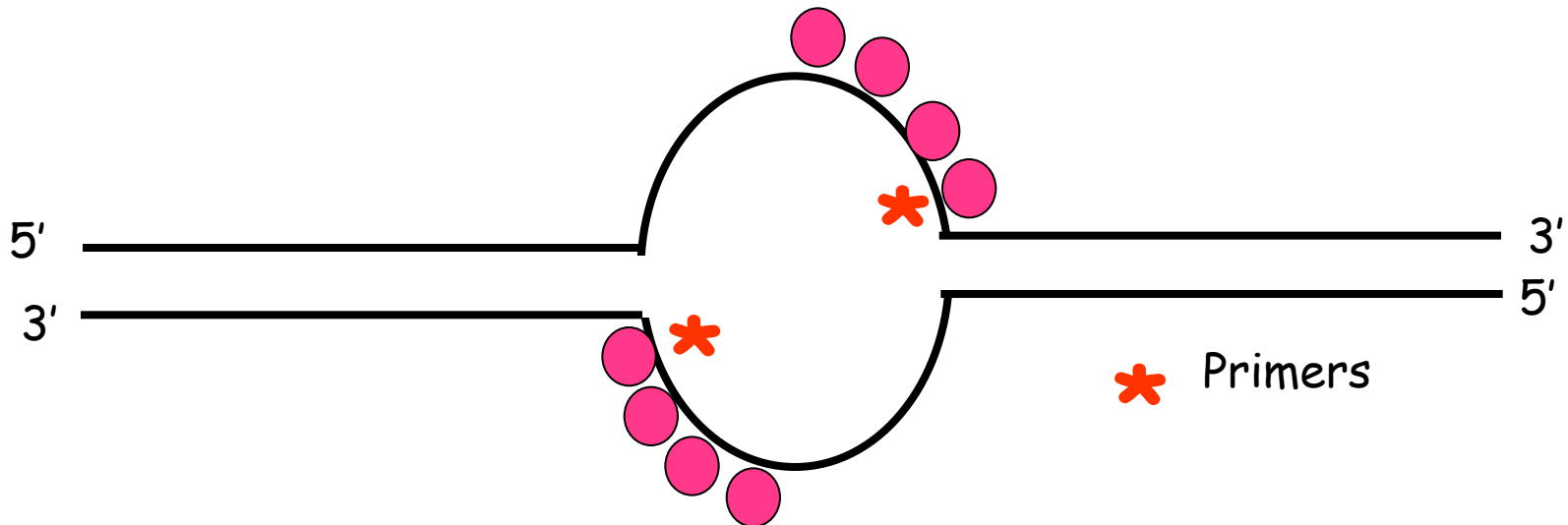
- - **Single-strand-binding proteins** help to stabilize the unwound strands.

- **Topoisomerase II** relieves strain on the double helix that is generated from unwinding.



Initiation of DNA replication

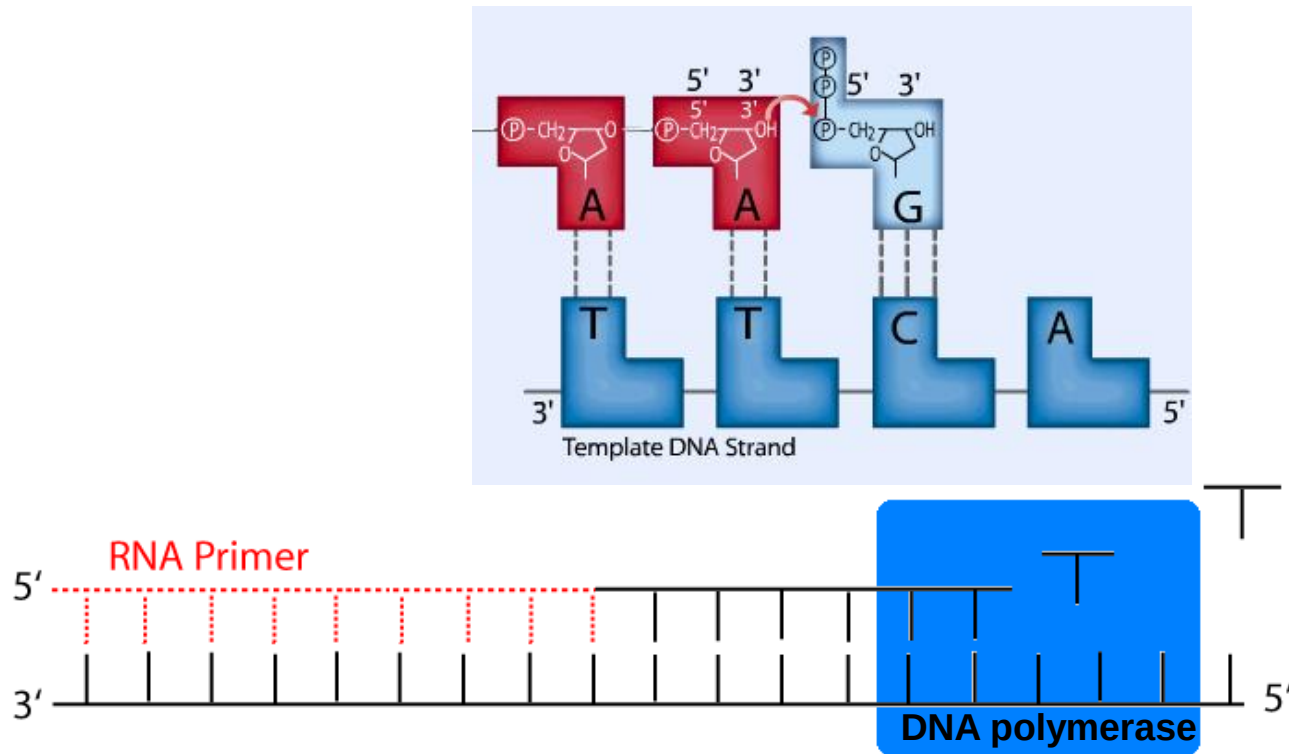
- Step 2 - binding of RNA primers
 - Primase adds short stretches of RNA primers
 - Purpose is to give DNA polymerase a 3'OH group from which to add new DNA nucleotides
 - Two primers are put down as the replication bubble opens



DNA Replication

Elongation

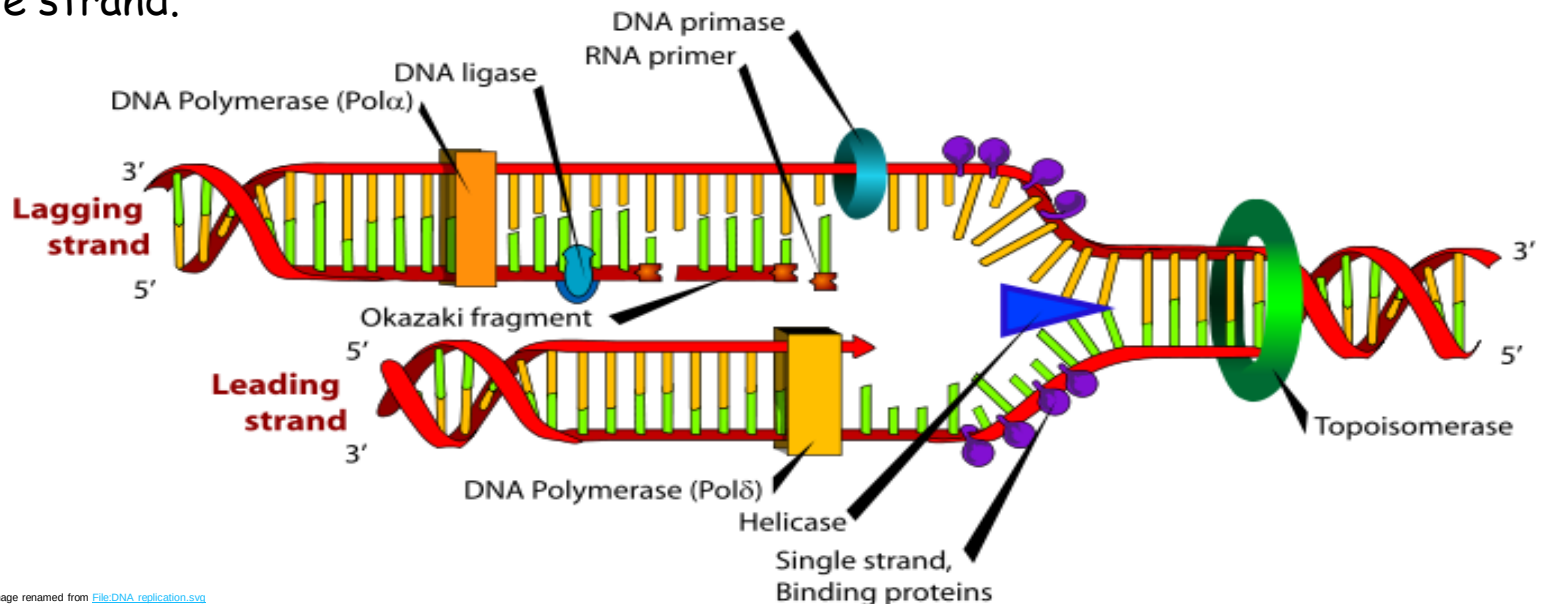
- **DNA polymerase** uses each strand as a template in the 3' to 5' direction to build a complementary strand in the 5' to 3' direction

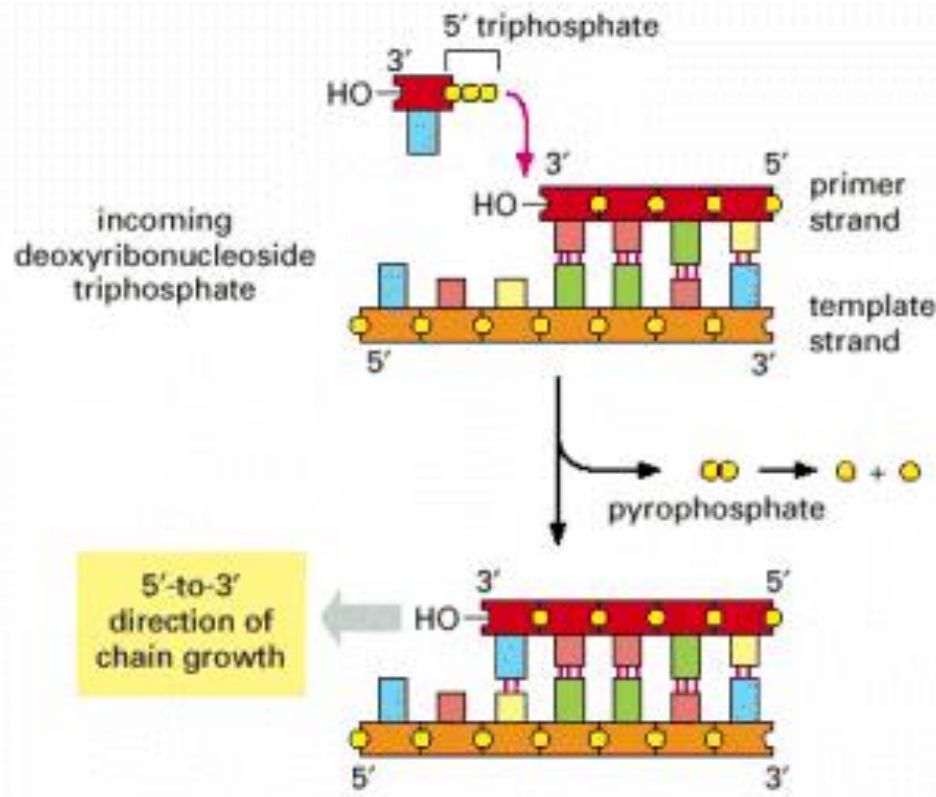


DNA Replication

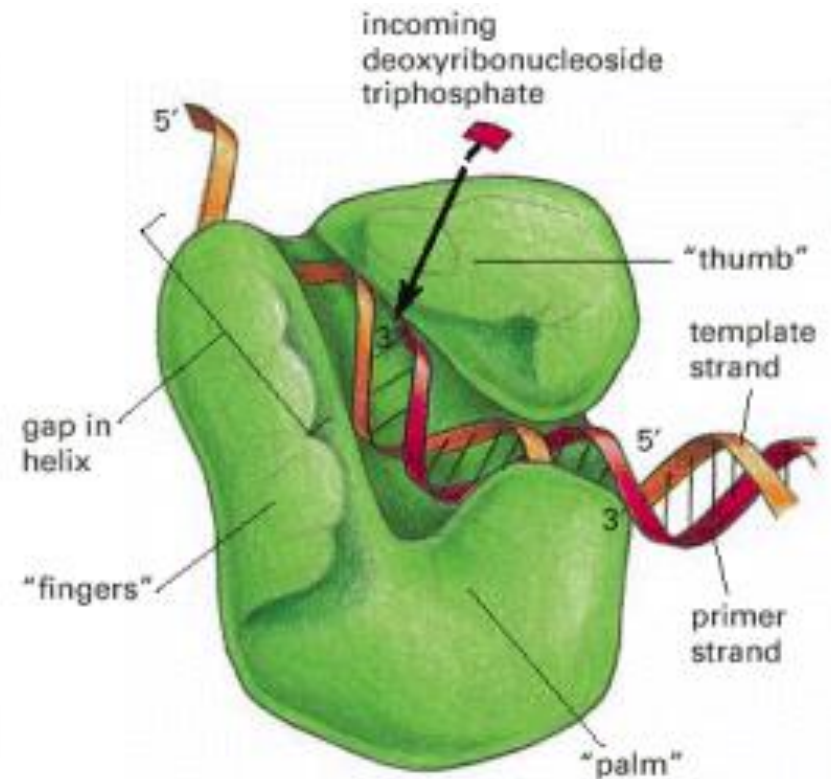
Elongation

- **DNA polymerase** uses each strand as a template in the 3' to 5' direction to build a complementary strand in the 5' to 3' direction
 - ✓ results in a **leading strand** and a **lagging strand**
- **DNA polymerase III** catalyzes the addition of new nucleotides to create a complementary strand to the parent strand. However, it can only attach new nucleotides to the free 3' hydroxyl end of a pre-existing chain of nucleotides.
- **DNA polymerase I** removes the primers and fills in the space by extending the neighboring DNA fragment. DNA ligase then joins the Okazaki fragments to create a complete strand.





(A)

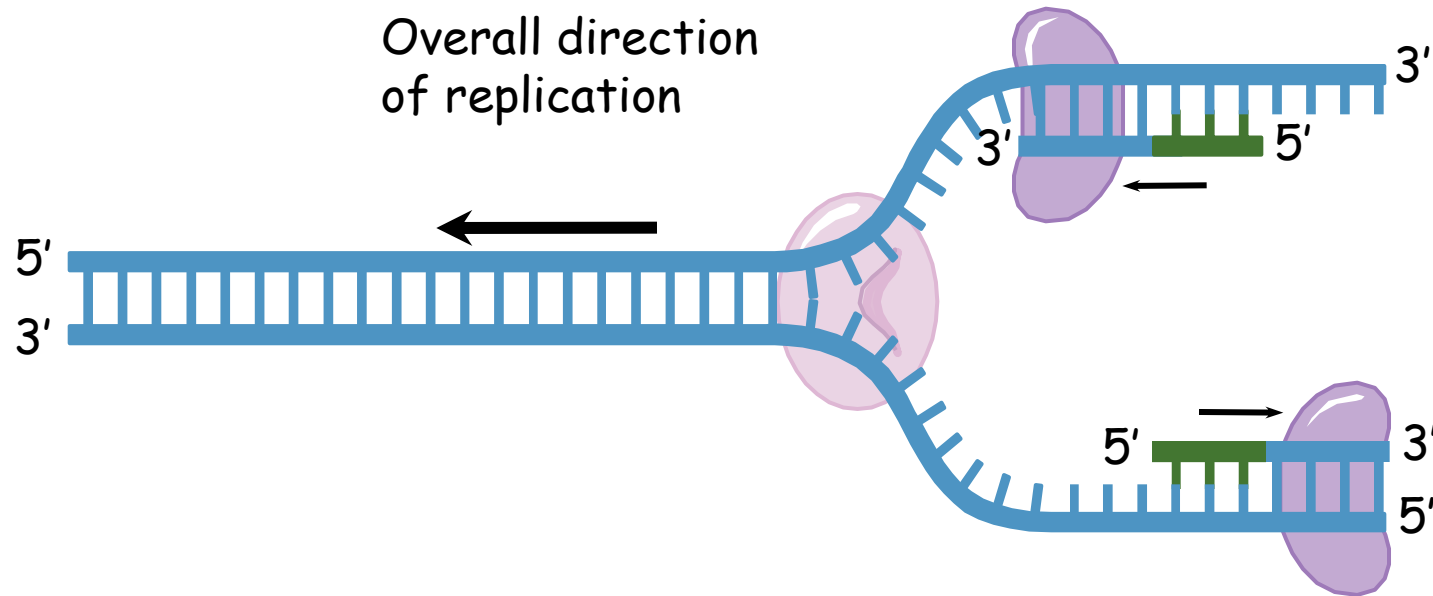


(B)

Role of DNA polymerases:

1. Polymerases catalyze the formation of phosphodiester bonds between the 3'-OH group of the deoxyribose on the last nucleotide and the 5'-phosphate of the dNTP precursor.
2. DNA polymerase finds the correct complement at each step in the process. 60-90 bases per second in humans.
3. The direction of synthesis is 5' to 3' only.

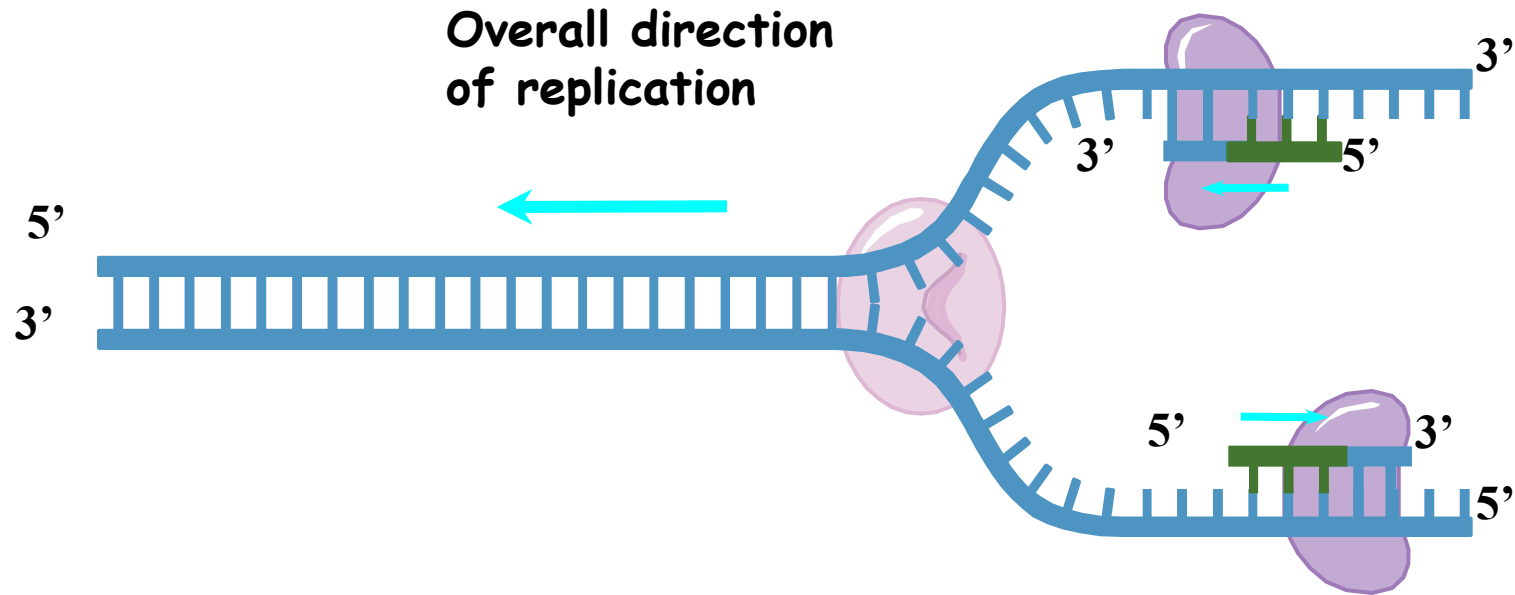
Replication



DNA polymerase enzyme adds DNA nucleotides to the RNA primer.

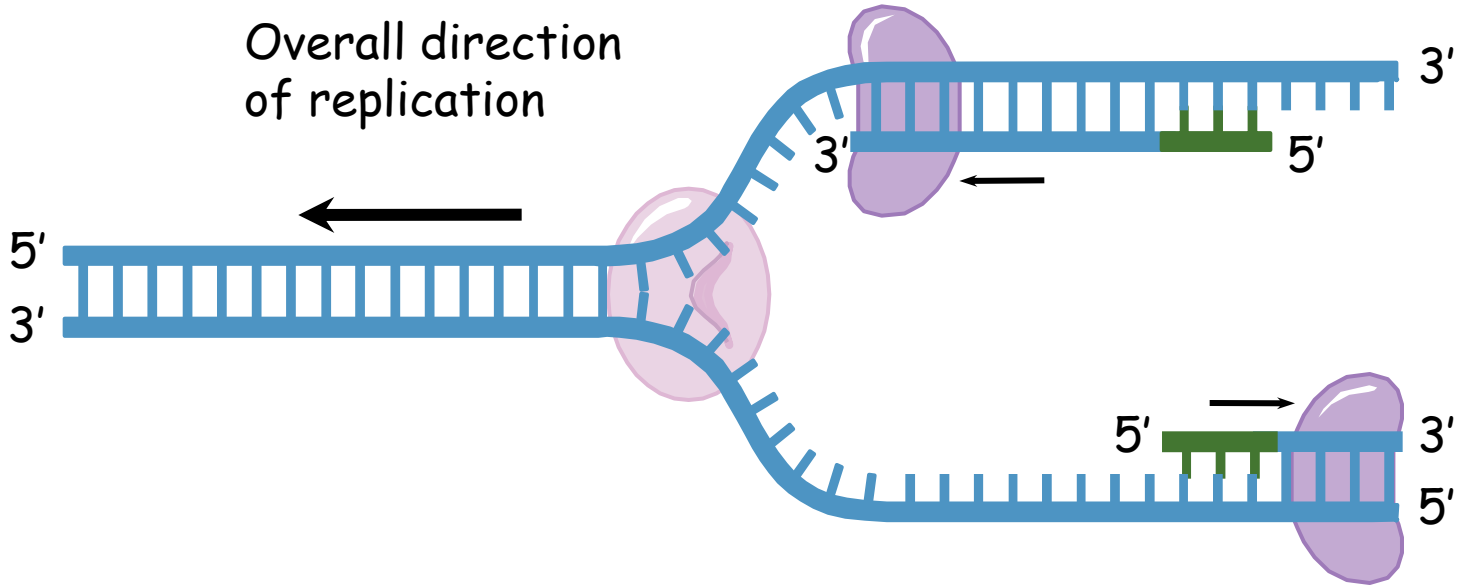
DNA polymerase proofreads bases added and replaces incorrect nucleotides.

Replication



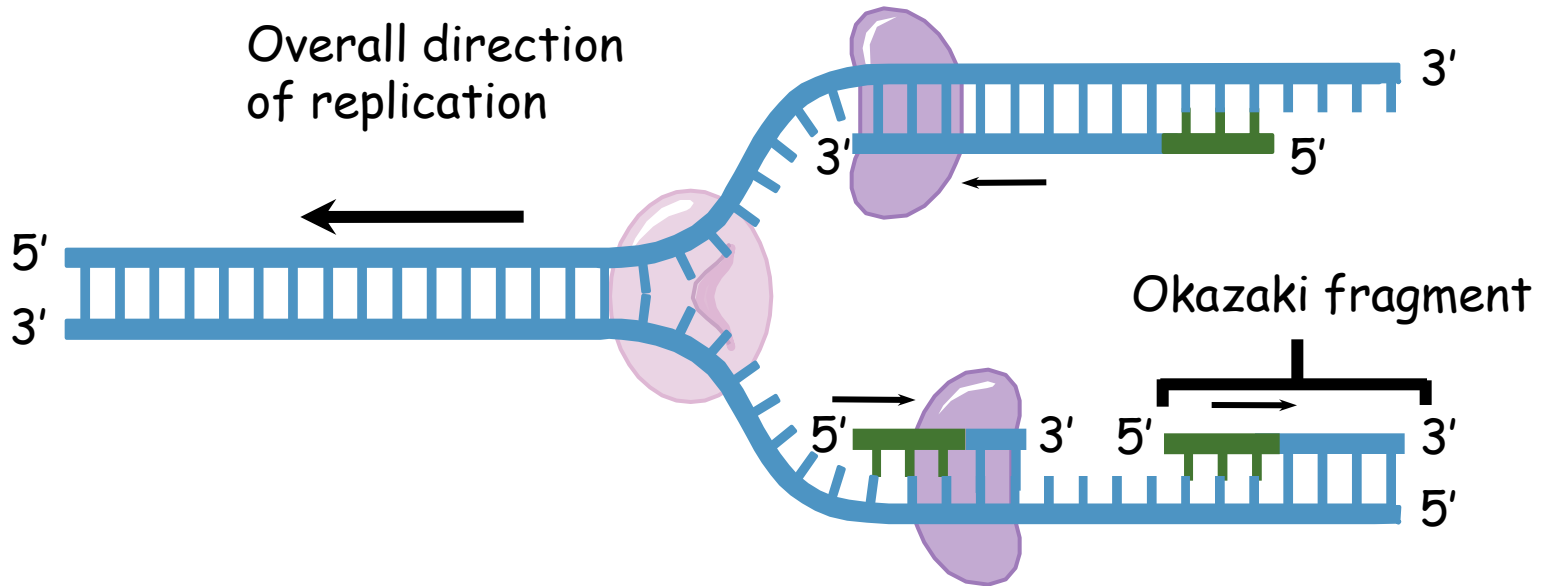
DNA polymerase enzyme adds DNA nucleotides to the RNA primer.

Replication



Leading strand synthesis continues in a 5' to 3' direction.

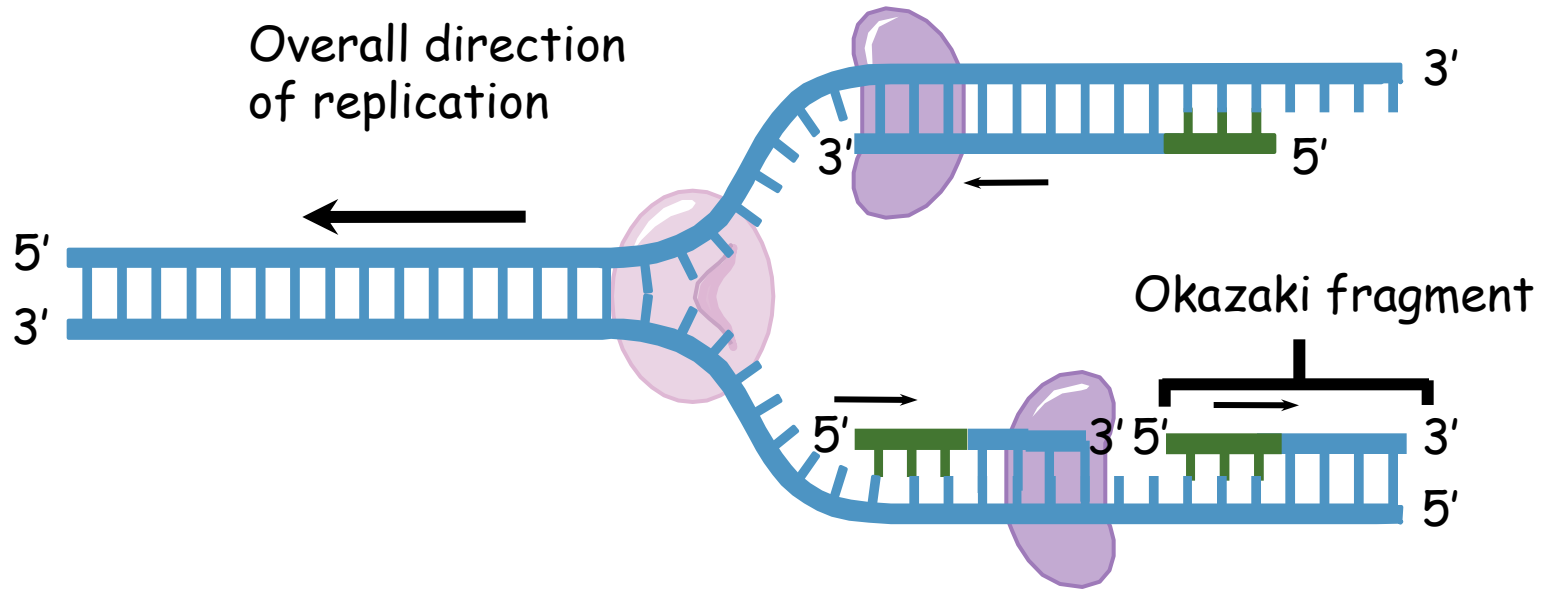
Replication



Leading strand synthesis continues in a 5' to 3' direction.

Discontinuous synthesis produces 5' to 3' DNA segments called Okazaki fragments.

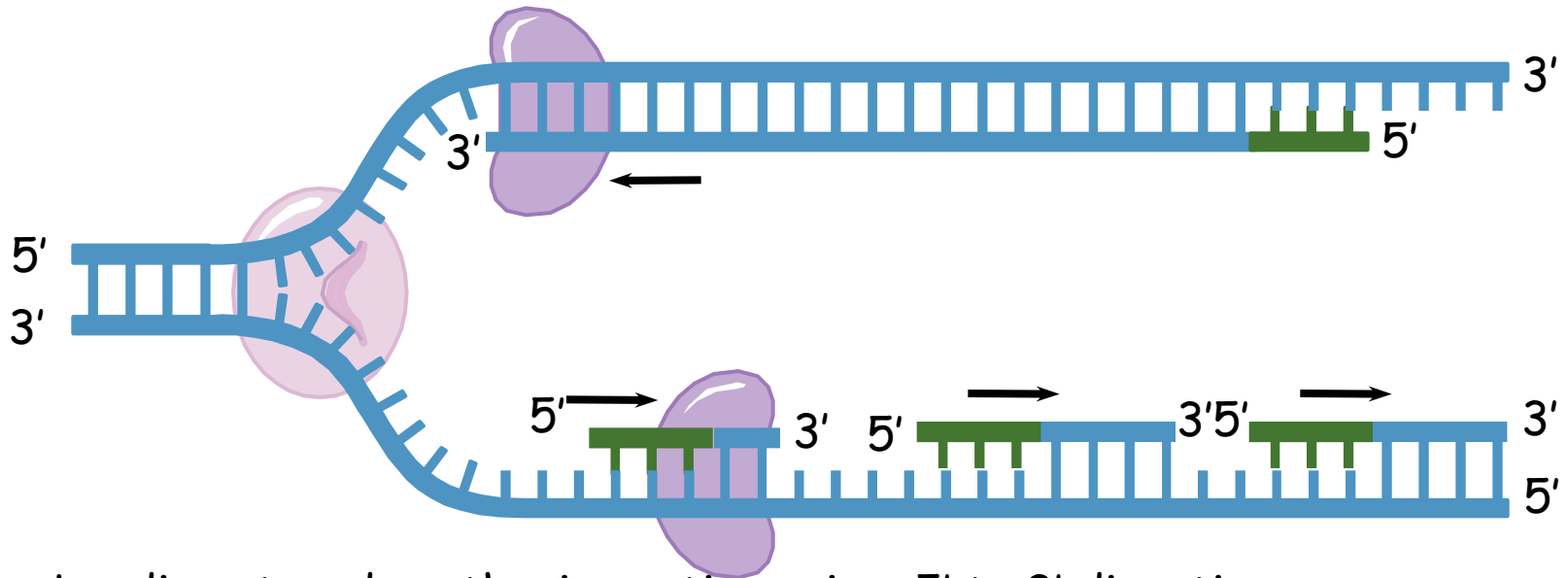
Replication



Leading strand synthesis continues in a 5' to 3' direction.

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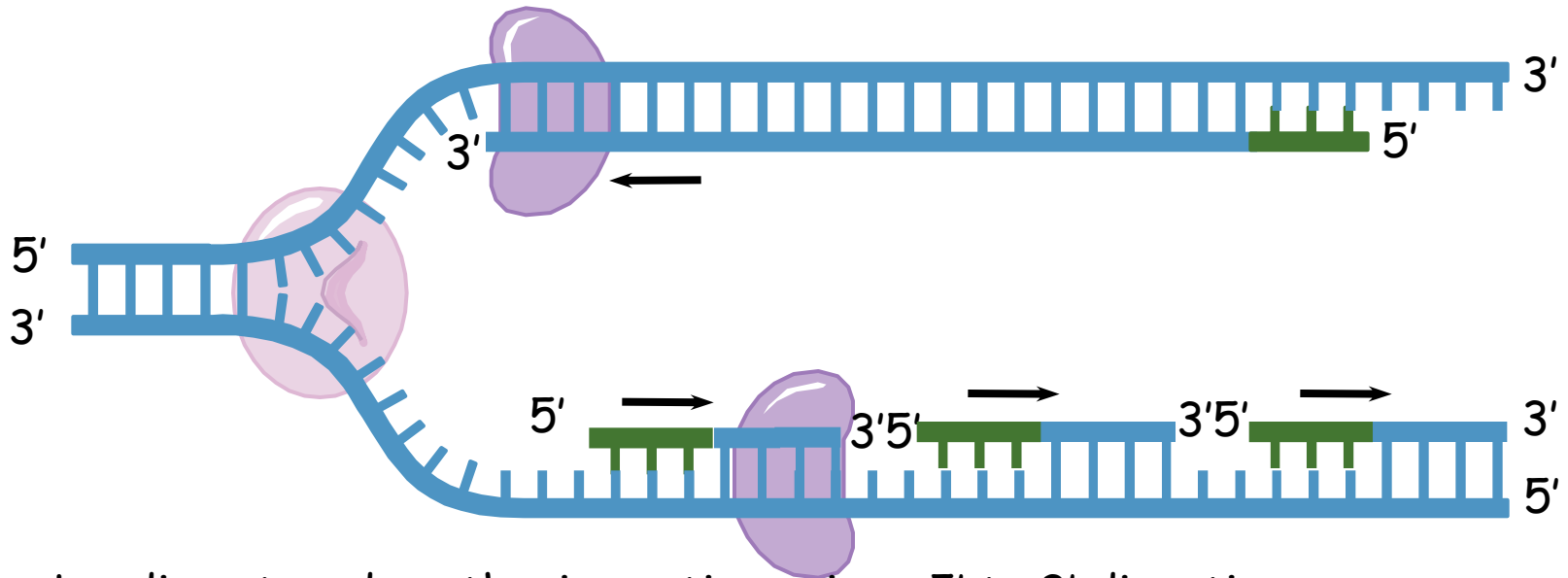
Replication



Leading strand synthesis continues in a 5' to 3' direction.

Discontinuous synthesis produces 5' to 3' DNA segments called Okazaki fragments.

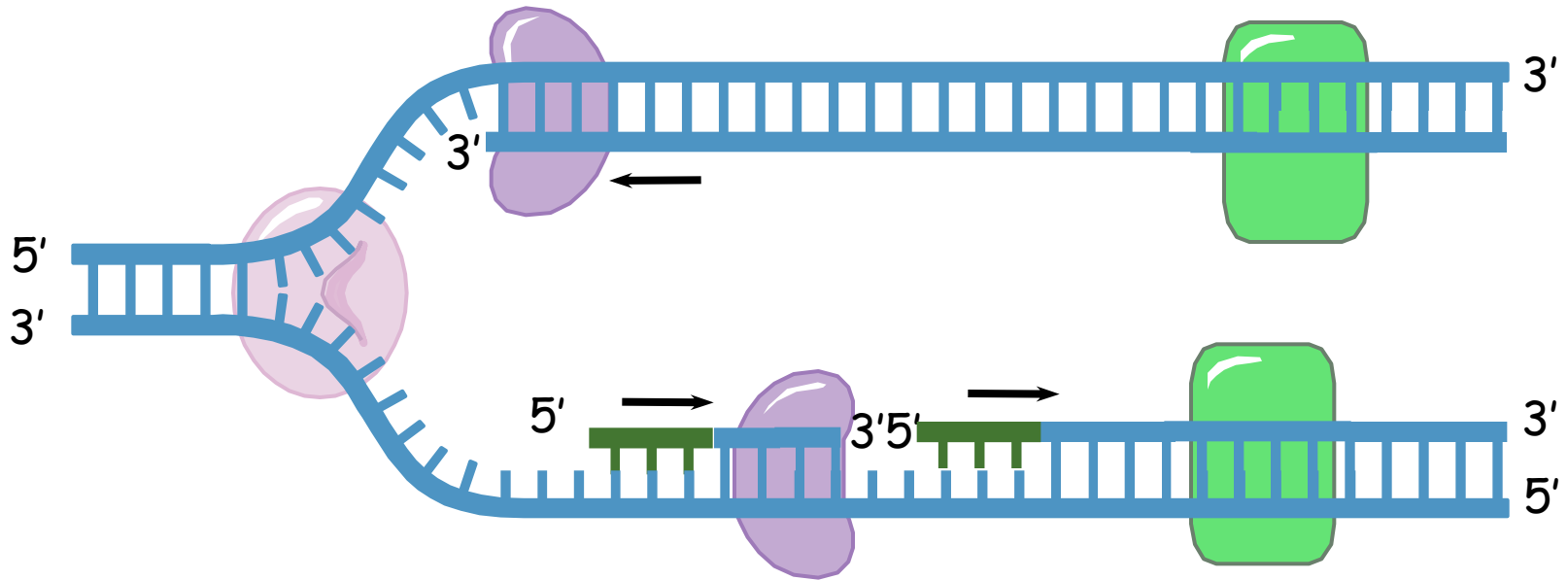
Replication



Leading strand synthesis continues in a 5' to 3' direction.

Discontinuous synthesis produces 5' to 3' DNA segments called Okazaki fragments.

Replication

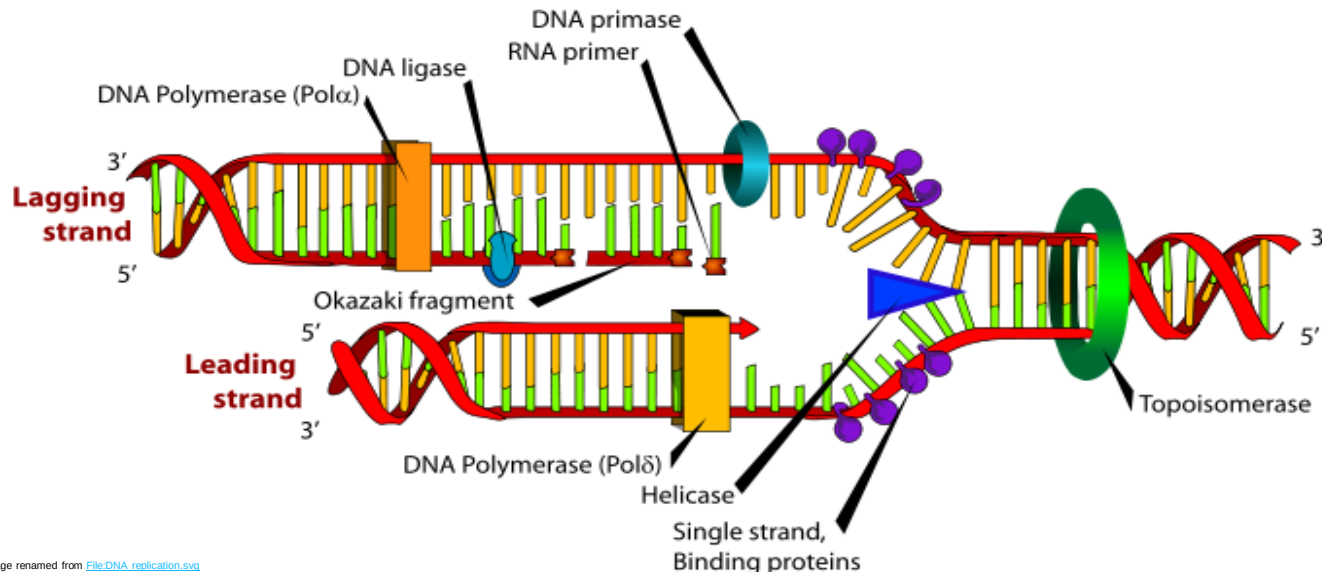


Polymerase activity of DNA polymerase I fills the gaps.
Ligase forms bonds between sugar-phosphate backbone.

DNA Replication

Leading Strand

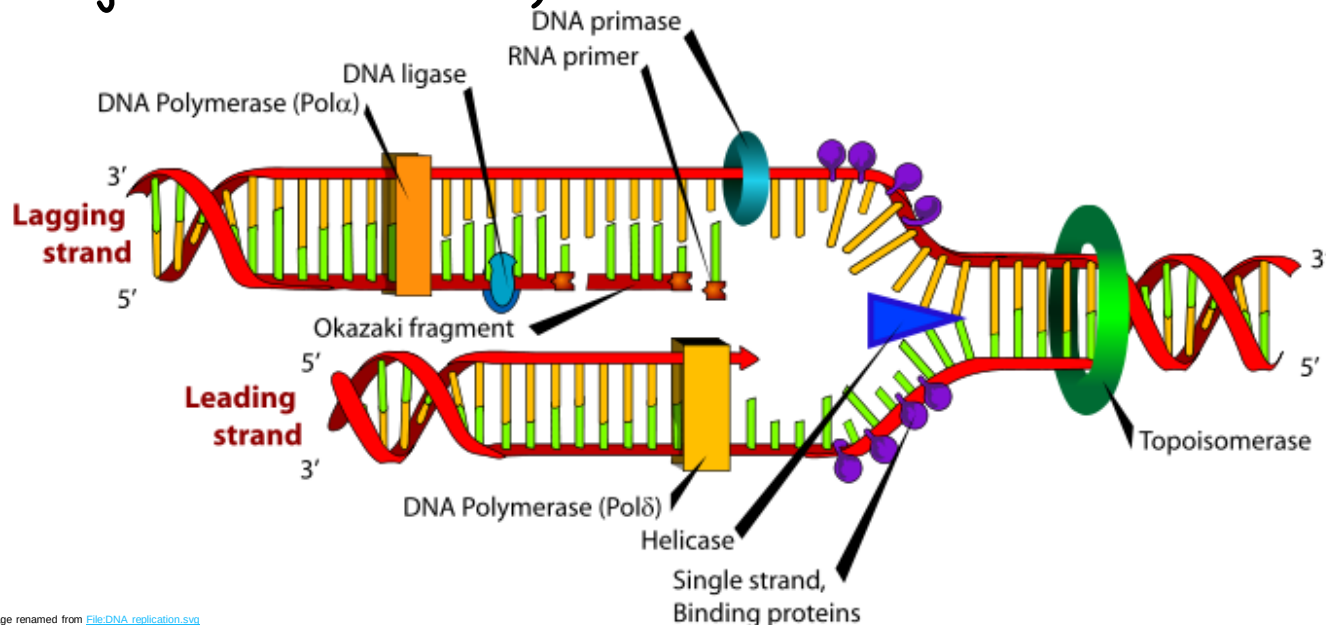
1. Topoisomerase unwinds DNA and then Helicase breaks H-bonds
2. DNA primase creates a single RNA primer to start the replication
3. DNA polymerase slides along the leading strand in the 3' to 5' direction synthesizing the matching strand in the 5' to 3' direction
4. The RNA primer is degraded by RNase H and replaced with DNA nucleotides by DNA polymerase, and then DNA ligase connects the fragment at the start of the new strand to the end of the new strand



DNA Replication

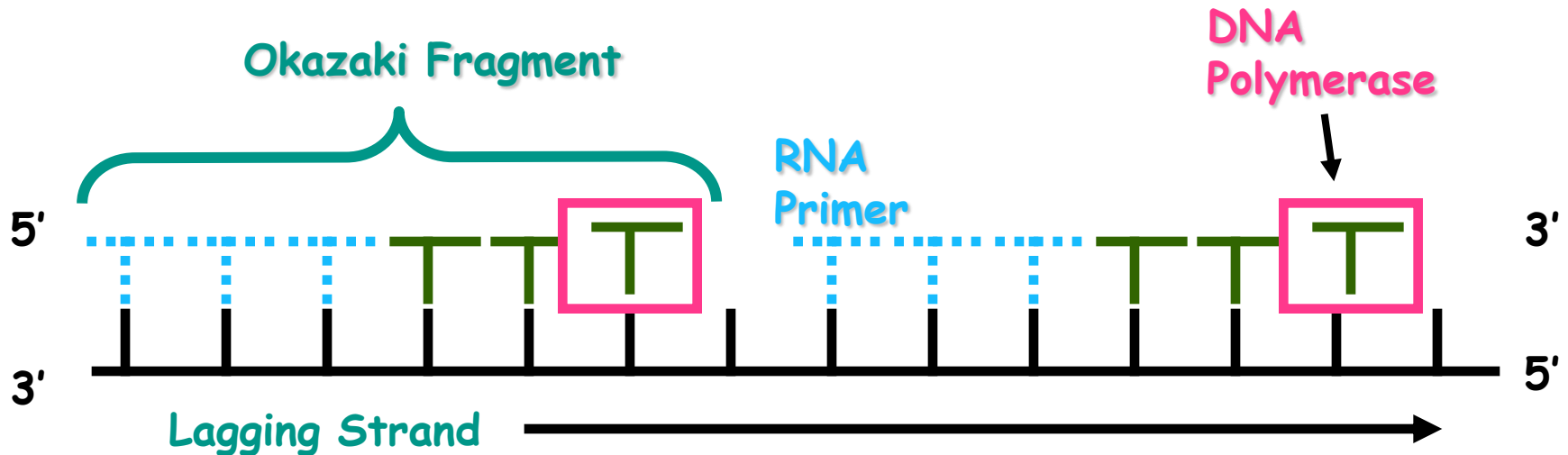
Lagging Strand

1. Topoisomerase unwinds DNA and then Helicase breaks H-bonds
2. DNA primase creates RNA primers in spaced intervals
3. DNA polymerase slides along the leading strand in the 3' to 5' direction synthesizing the matching Okazaki fragments in the 5' to 3' direction
4. The RNA primers are degraded by RNase H and replaced with DNA nucleotides by DNA polymerase
5. DNA ligase connects the Okazaki fragments to one another (covalently bonds the phosphate in one nucleotide to the deoxyribose of the adjacent nucleotide)



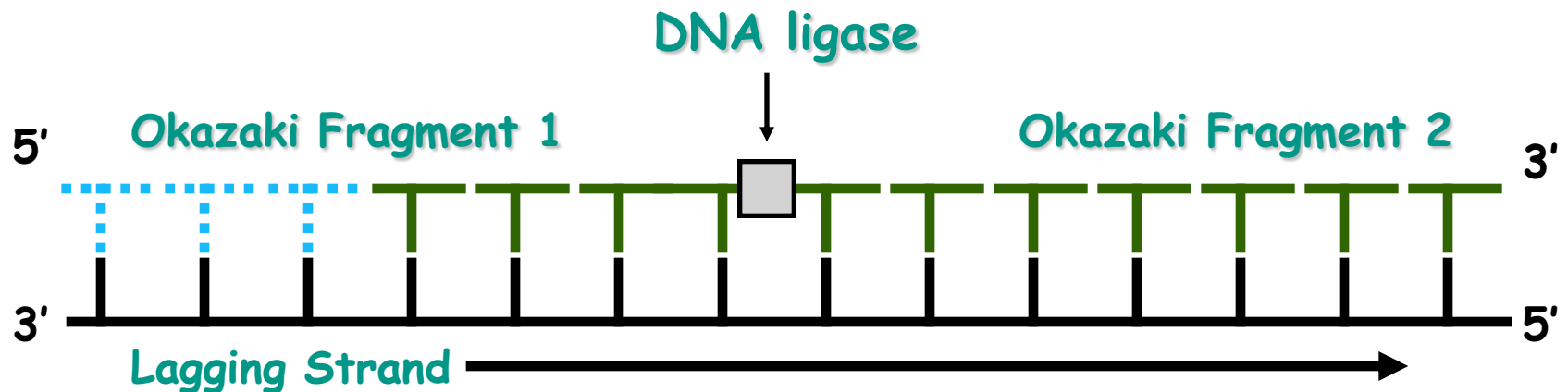
Lagging Strand Segments

- **Okazaki Fragments** - series of short segments on the lagging strand
- Must be joined together by an **enzyme**



Joining of Okazaki Fragments

- The enzyme **Ligase** joins the Okazaki fragments together to make one strand



- Leading strand

synthesized 5' to 3' in the direction of the replication fork movement.

continuous

requires a single RNA primer

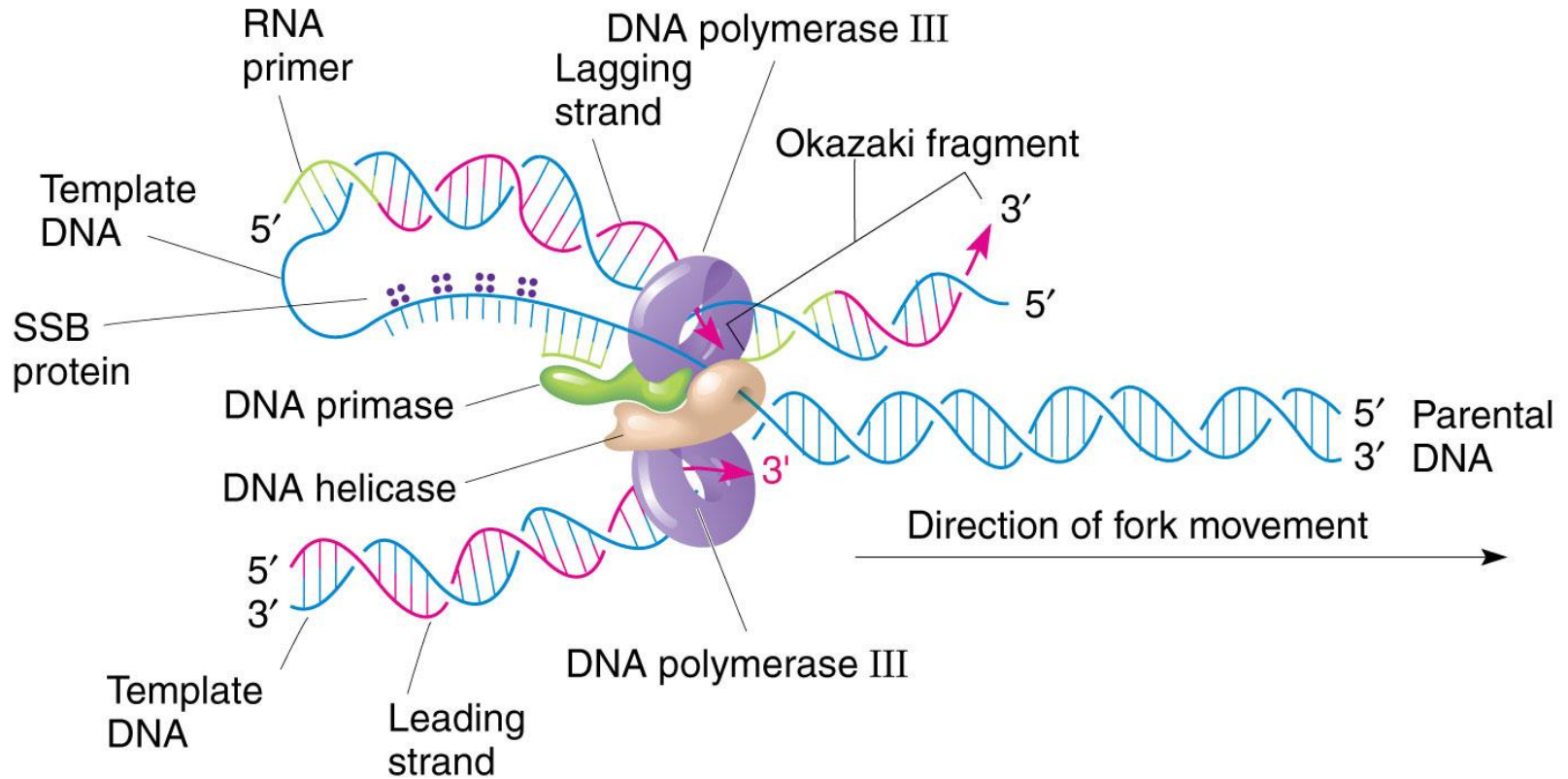
- Lagging strand

synthesized 5' to 3' in the opposite direction.

discontinuous (i.e., not continuous)

requires many RNA primers , DNA is synthesized in short fragments.

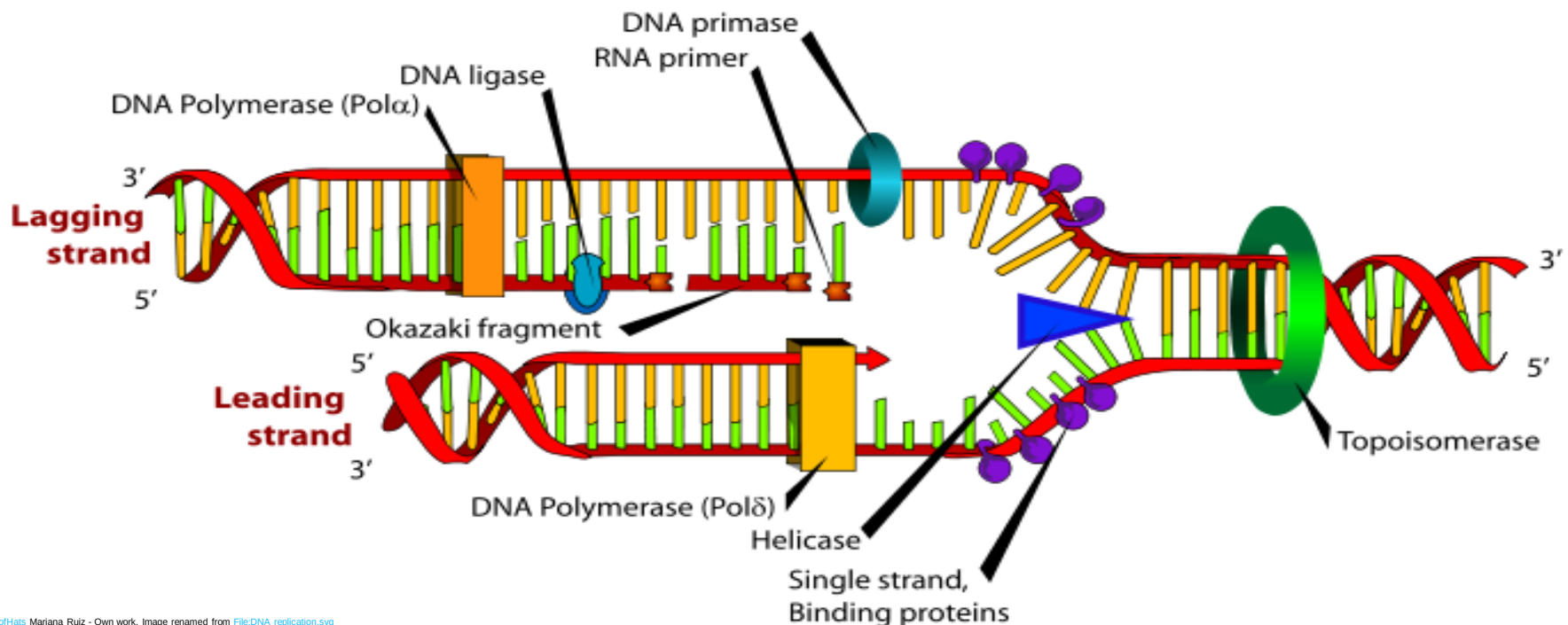
Model of DNA Replication



A General Model for DNA Replication

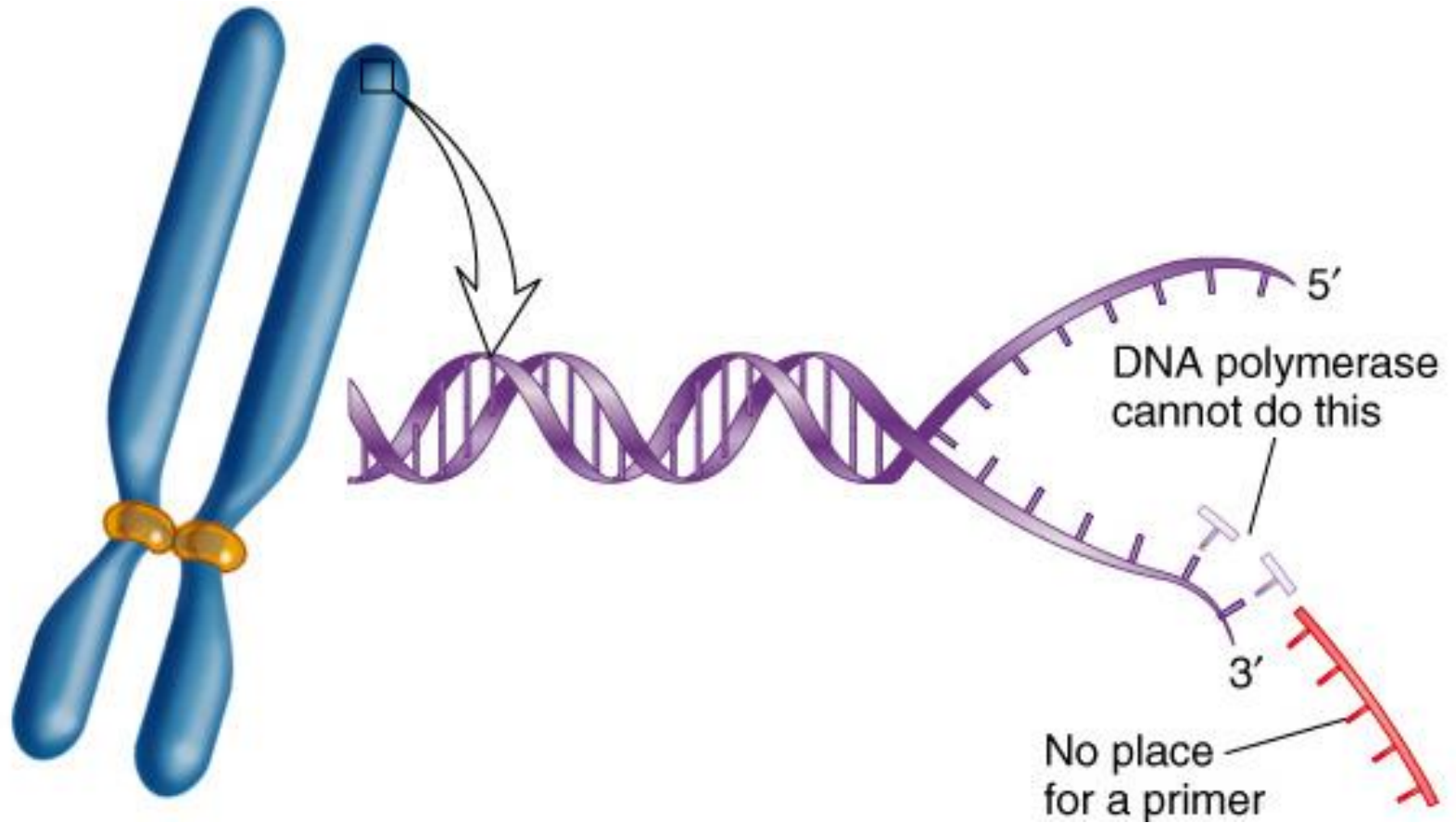
1. The DNA molecule is unwound and prepared for synthesis by the action of DNA gyrase, DNA helicase and the single-stranded DNA binding proteins.
2. A free 3'OH group is required for replication, but when the two chains separate no group of that exists in nature therefore RNA primers are synthesized, and the free 3'OH of the primer is used to begin replication.

3. The replication fork moves in one direction, but DNA replication only goes in the 5' to 3' direction. This paradox is resolved by the use of Okazaki fragments. These are short, discontinuous replication products that are produced off the lagging strand. This is in comparison to the continuous strand that is made off the leading strand.



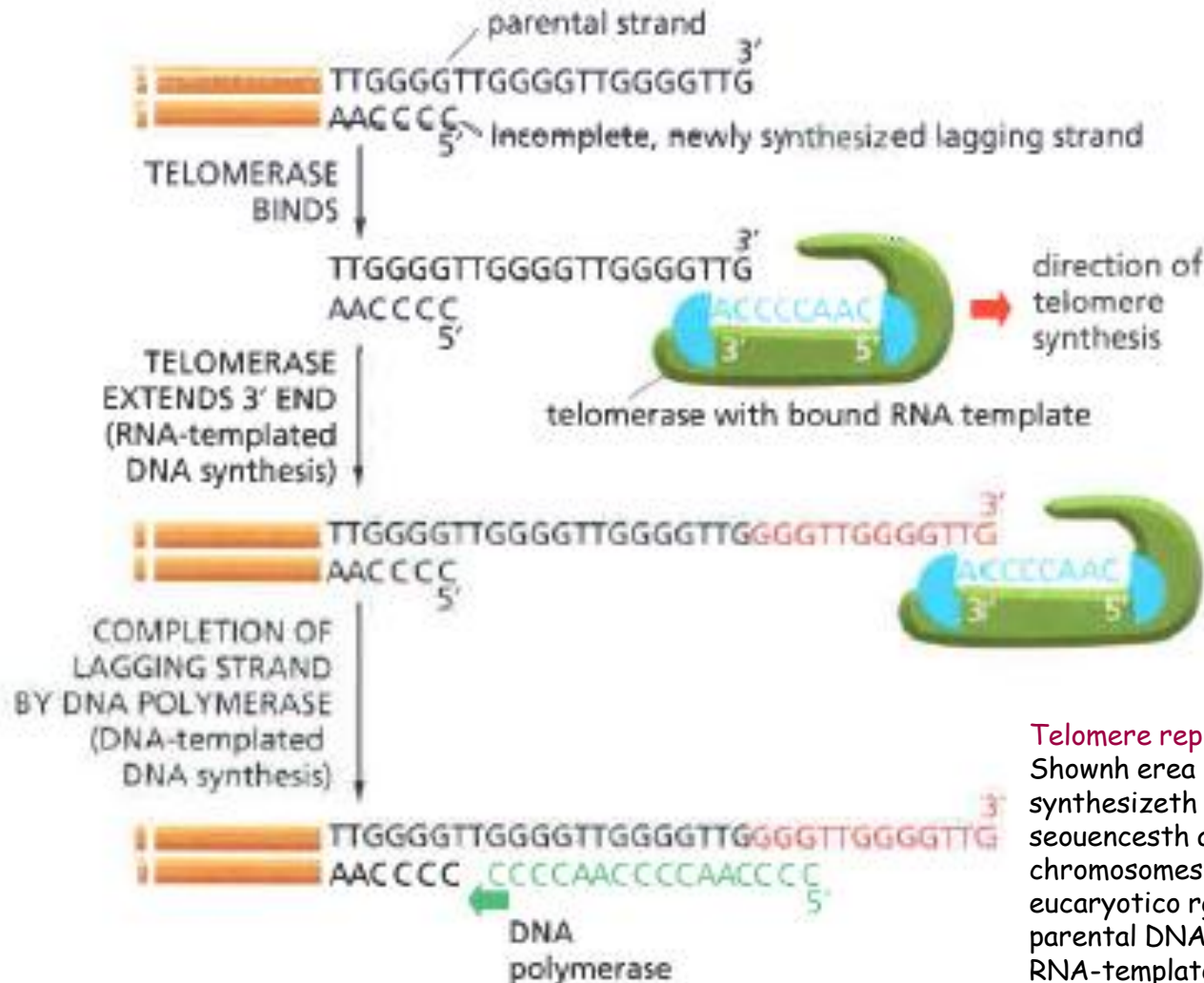
4. The final product does not have RNA stretches in it. These are removed by the 5' to 3' exonuclease action of Polymerase I.
5. The final product does not have any gaps in the DNA that result from the removal of the RNA primer. These are filled in by the 5' to 3' polymerase action of DNA Polymerase I.
6. DNA polymerase does not have the ability to form the final bond. This is done by the enzyme DNA ligase.

- DNA polymerases can only synthesize DNA only in the 5' to 3' direction and cannot initiate DNA synthesis
- These two features pose a problem at the 3' end of linear chromosomes



Problem at ends of eukaryotic linear Chromosomes

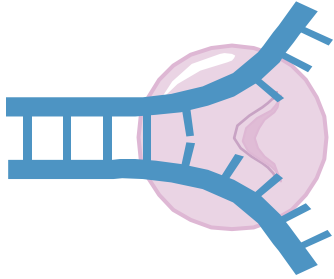
- If this problem is not solved
 - The linear chromosome becomes progressively shorter with each round of DNA replication
- The cell solves this problem by adding DNA sequences to the ends of chromosome:
telomeres
 - Small repeated sequences (100-1000's)
- Catalyzed by the enzyme telomerase
- Telomerase contains **protein** and **RNA**
 - The RNA functions as the template
 - complementary to the DNA sequence found in the telomeric repeat
 - This allows the telomerase to bind to the 3' overhang



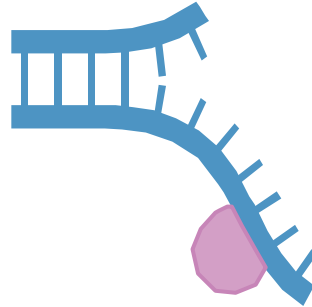
Telomere replication.

Shown here are the reactions that synthesize the repeating G-rich sequence at the ends of the chromosomes (telomeres) of diverse eucaryotic organisms. The 3' end of the parental DNA strand is extended by RNA-templated DNA synthesis; this allows the incomplete daughter DNA strand that is paired with it to be extended in its 5' direction. This incomplete lagging strand is presumed to be completed by DNA polymerase, which carries a DNA primase so one of its subunits.

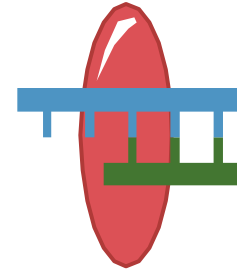
Enzymes in DNA replication



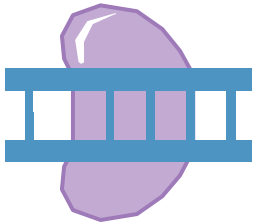
Helicase unwinds
parental double helix



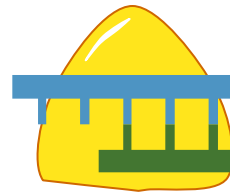
Binding proteins
stabilize separate
strands



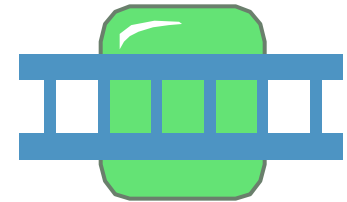
Primase adds
short primer
to template strand



DNA polymerase III
binds nucleotides
to form new strands

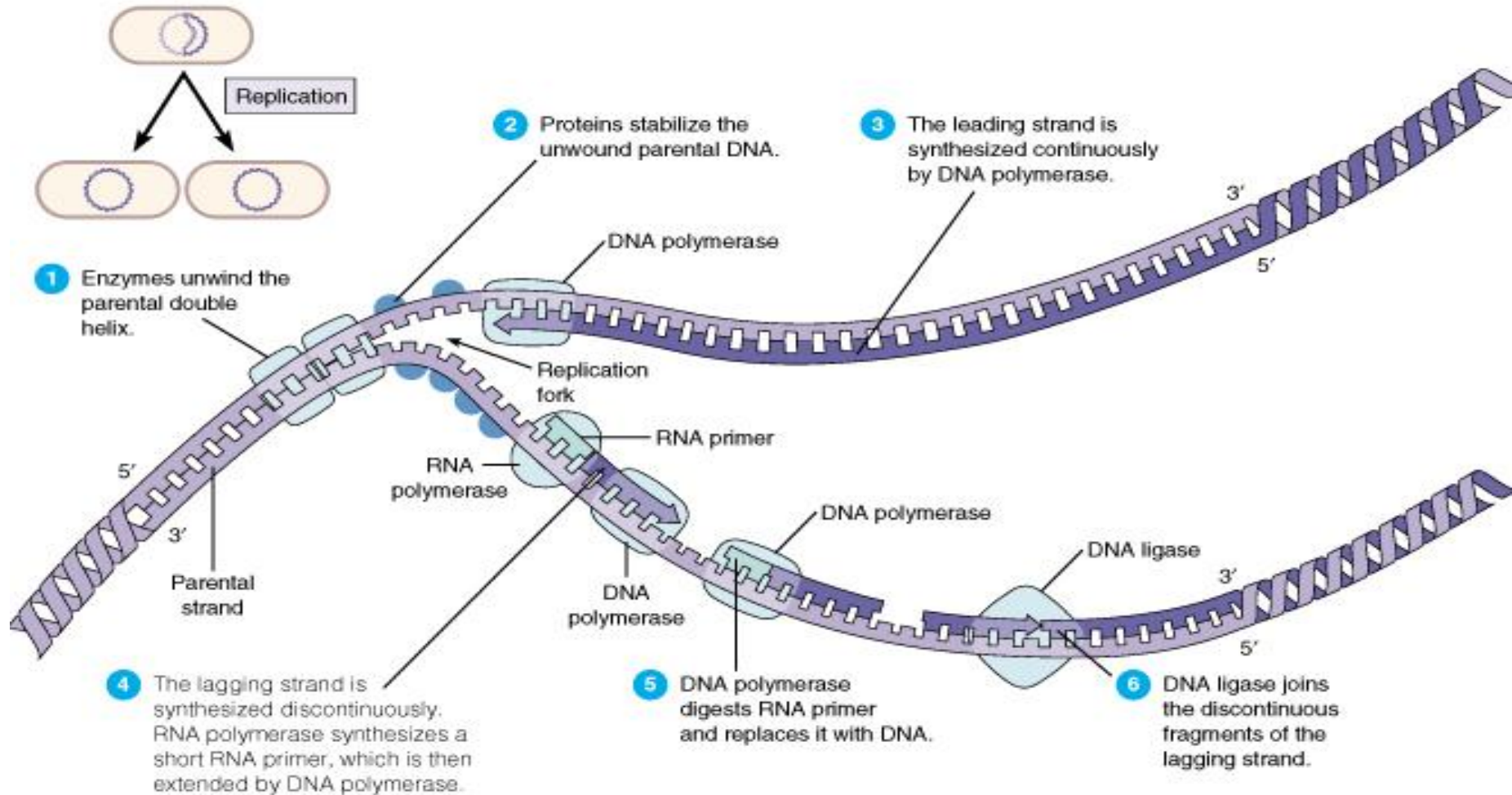


DNA polymerase I
(Exonuclease) removes
RNA primer and inserts
the correct bases



Ligase joins Okazaki
fragments and seals
other nicks in sugar-
phosphate backbone

Replication Fork Overview



Risks To DNA Replication

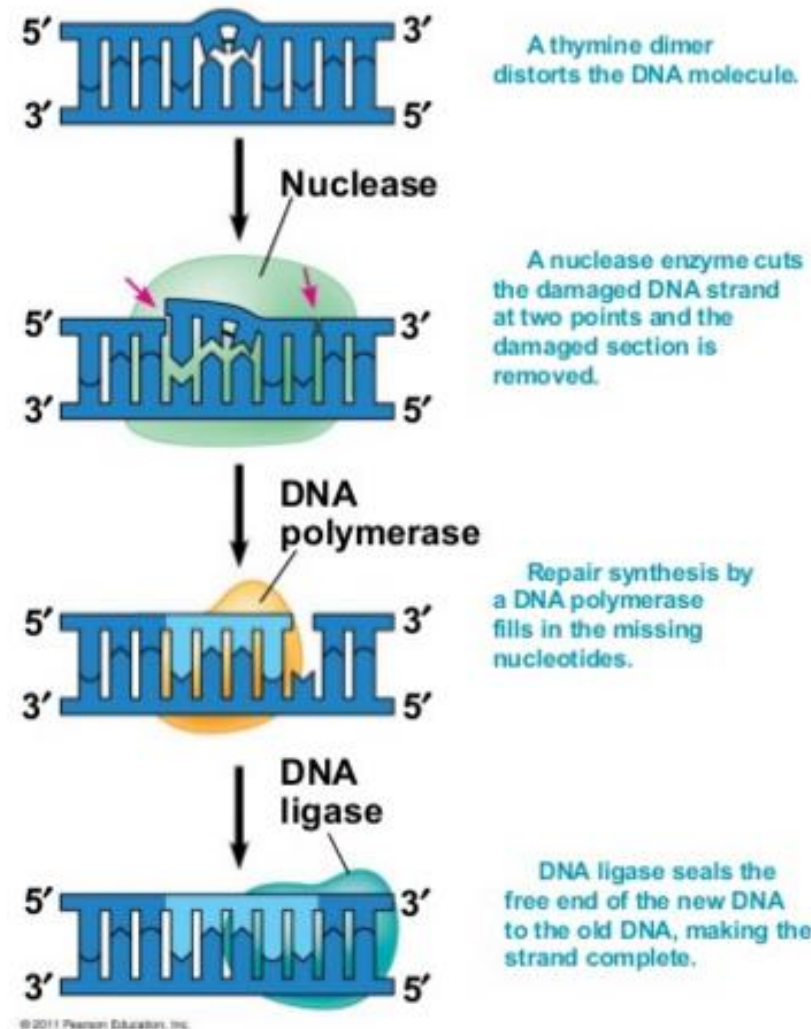
- DNA polymerase inserts the incorrect base once in every 100,000 bases
 - Error rate of 1×10^{-5}
 - At this rate your genome would be riddle with mutations
- But as it turns out DNA polymerase can proofread

Mistakes during Replication

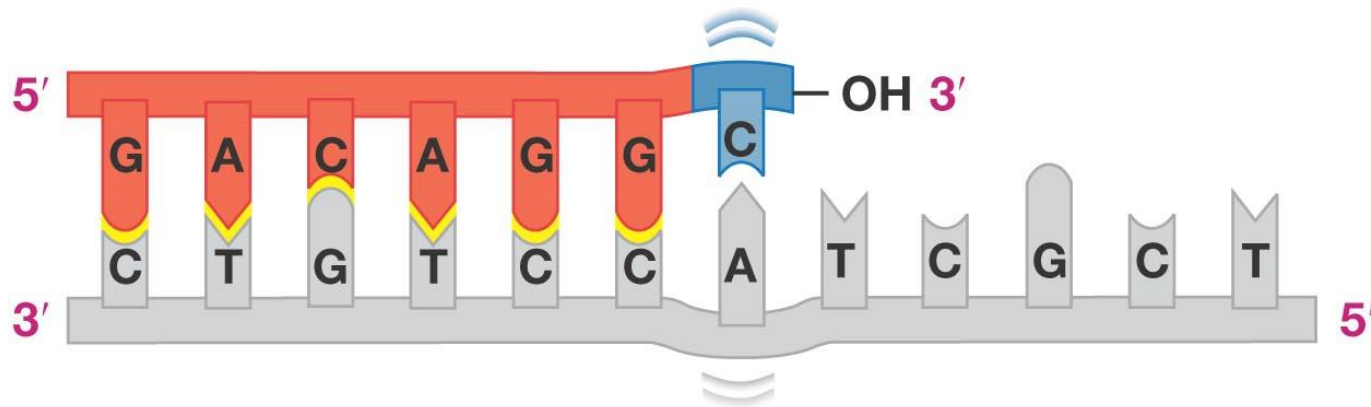
- Base pairing rules must be maintained
 - Mistake = genome mutation, may have consequence on daughter cells
- Only correct pairings fit in the polymerase active site
- If wrong nucleotide is included
 - Polymerase uses its **proofreading** ability to cleave the phosphodiester bond of improper nucleotide
 - Activity $3' \rightarrow 5'$
 - And then adds correct nucleotide and proceeds down the chain again in the $5' \rightarrow 3'$ direction

Proofreading and Repairing DNA

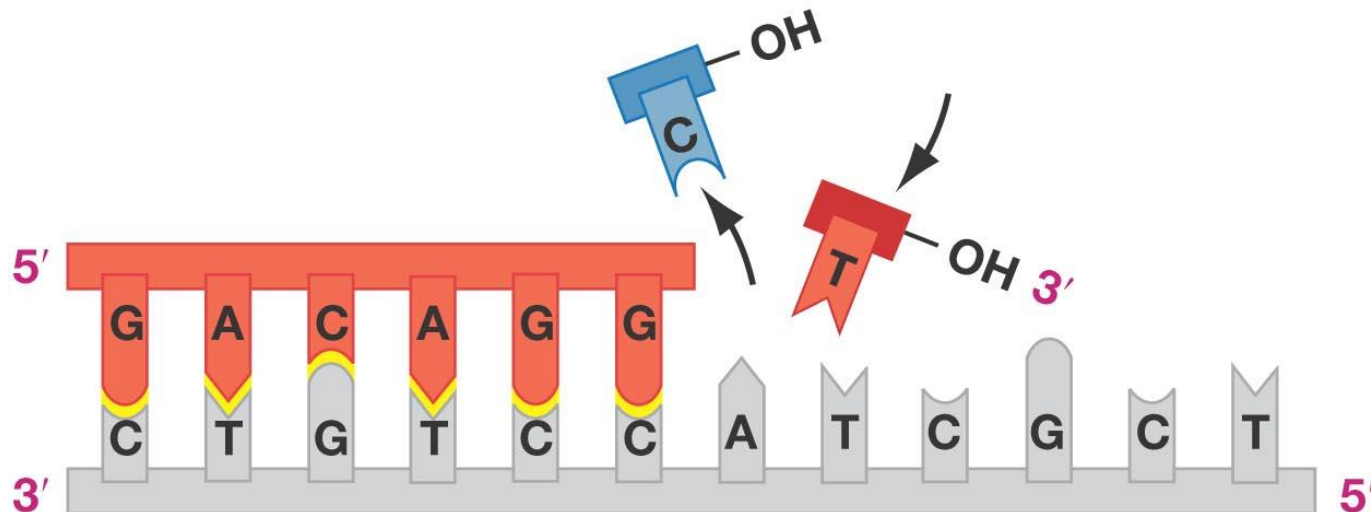
- DNA polymerases proofread newly made DNA, replacing any incorrect nucleotides
- Mismatch repair: 'wrong' inserted base can be removed
- Excision repair: DNA may be damaged by chemicals, radiation, etc. Mechanism to cut out and replace with correct bases



(a) Mismatched bases



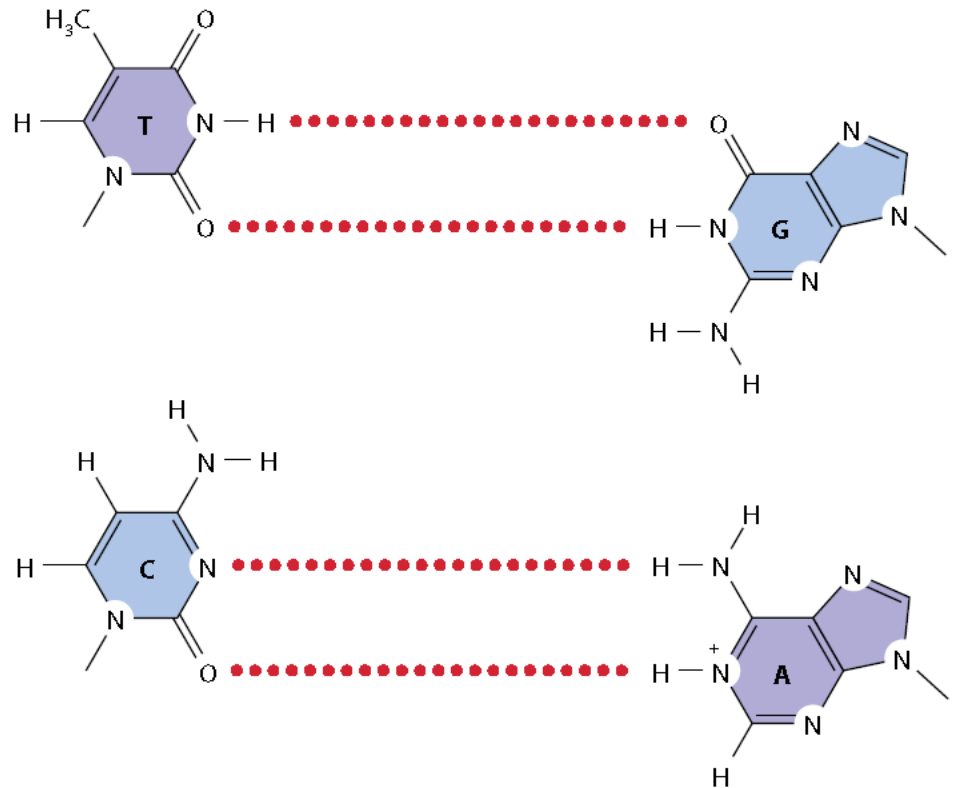
(b) DNA polymerase III can repair mismatches.



Errors During DNA Replication

A human cell can copy its entire DNA in a few hours, with an error rate of about one per 1 billion nucleotide pairs.

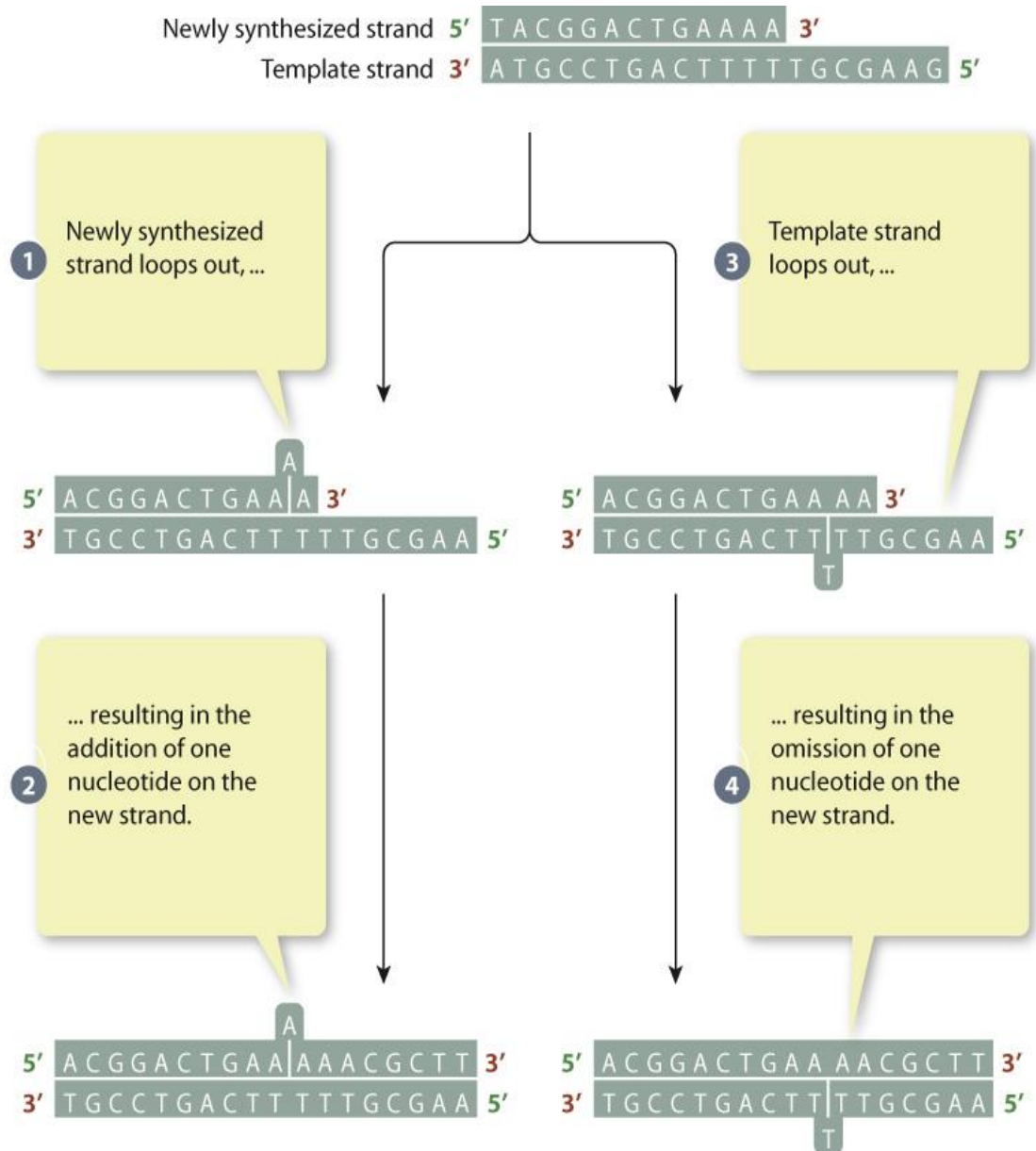
- Errors naturally occur during replication.
- Mismatching of bases and strand slippage are two types of errors that cause either additions or omissions of nucleotides.



Incorrect pairing (mismatching) of bases is thought to occur as a result of flexibility in DNA structure.

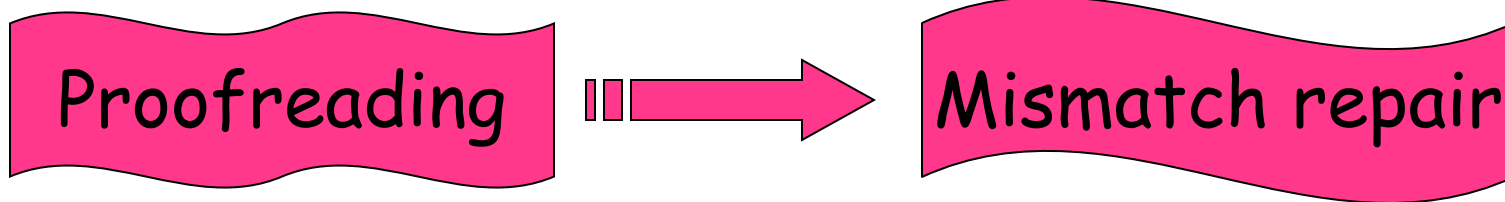
Errors During DNA Replication

Strand slippage during DNA replication can cause the addition or omission of nucleotides in newly synthesized strands, which represent errors.



Replication errors and their repair

Repair:



Factors Influencing the Rate of Spontaneous Mutations

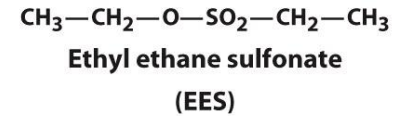
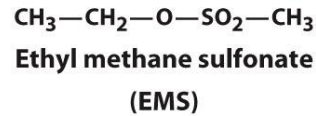
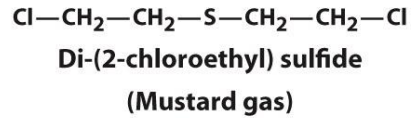
- Accuracy of the DNA replication machinery
- Efficiency of the mechanisms for the repair of damaged DNA
- Degree of exposure to mutagenic agents in the environment

Types of Chemical Mutagens

- Chemicals that are mutagenic to both replicating and nonreplicating DNA (e.g., alkylating agents and nitrous acid)
- Chemicals that are mutagenic only to replicating DNA (e.g., base analogs and acridine dyes)

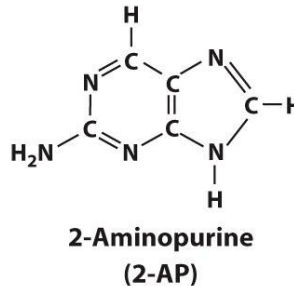
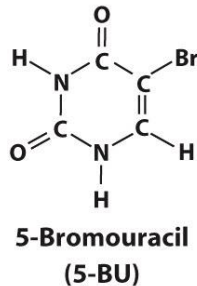
Chemical Mutagens

Alkylating agents



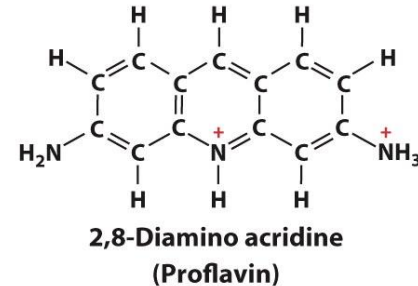
(a)

Base analogs

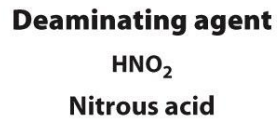


(b)

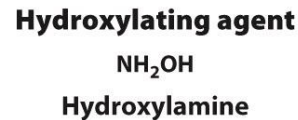
Acridines



(c)



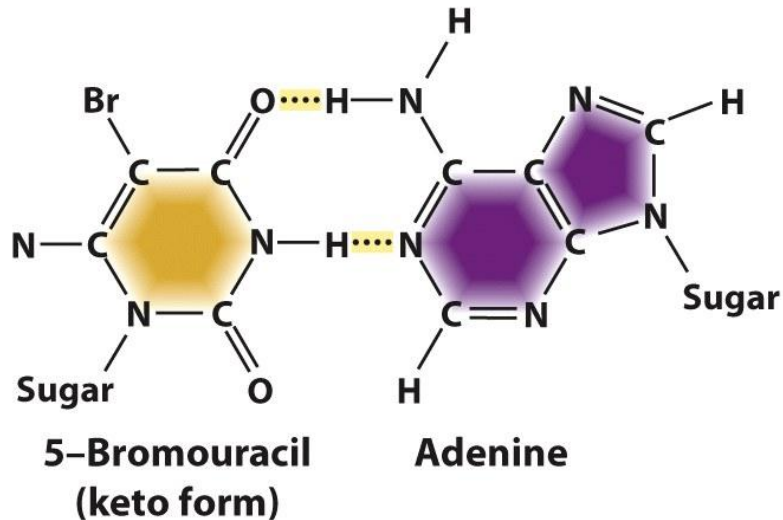
(d)



(e)

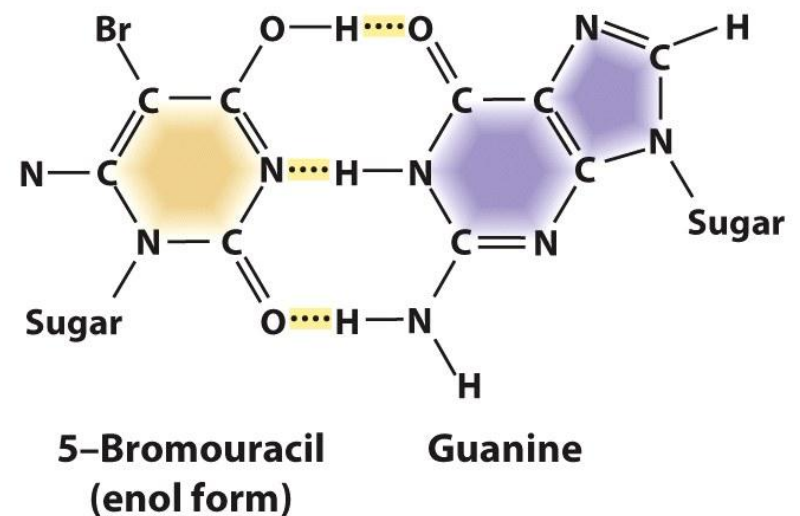
A Base Analog: 5-Bromouracil

5-Bromouracil : adenine base pair.



(a)

5-Bromouracil : guanine base pair.



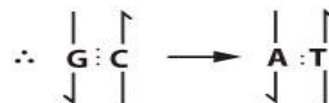
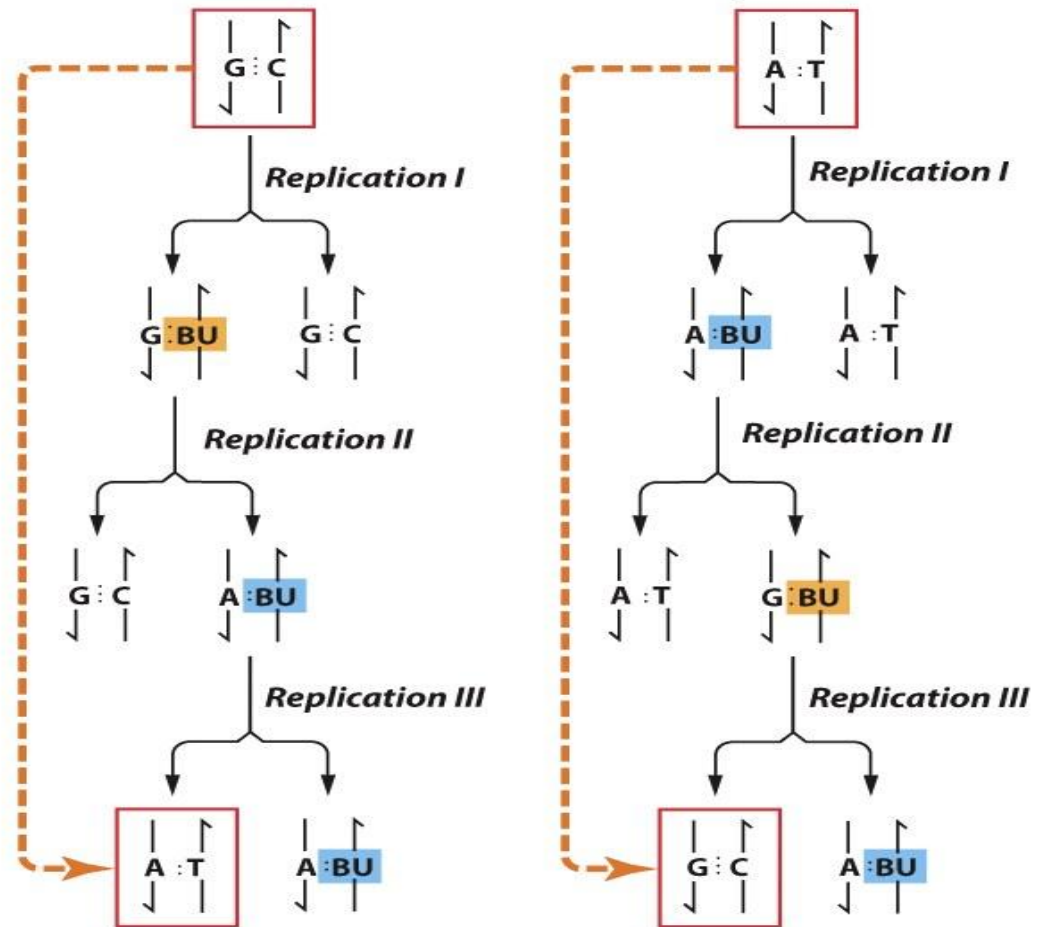
(b)

Mutagenic Effects of 5-Bromouracil

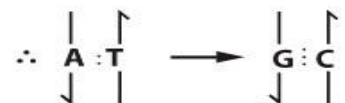
Effect of enol form of 5-bromouracil during:

Incorporation

Replication after incorporation



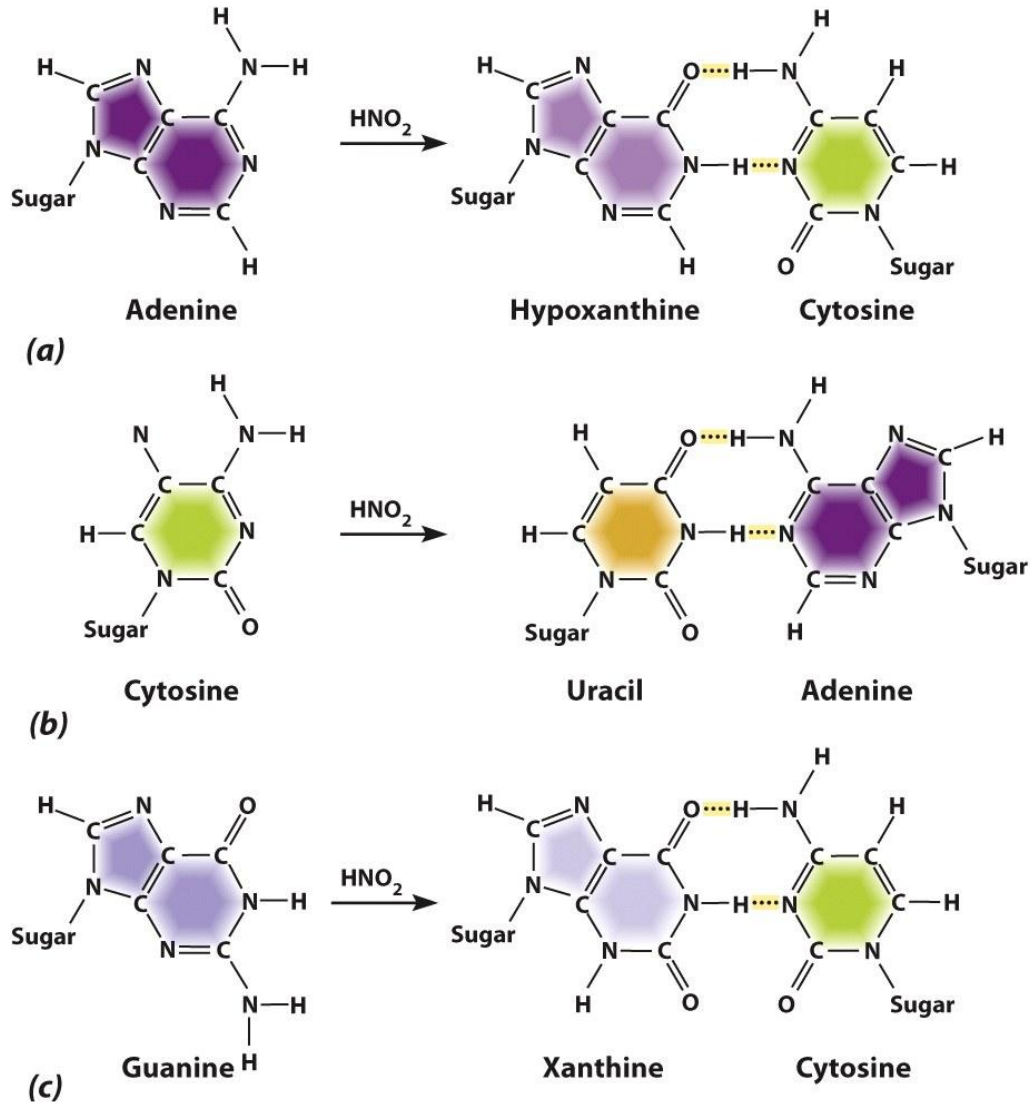
(a)



(b)



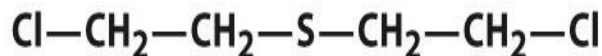
Nitrous Acid Causes Oxidative Deamination of Bases



Alkylating Agents

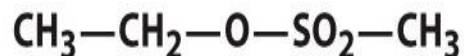
- chemicals that donate alkyl groups to other molecules.
- induce transitions, transversions, frameshifts, and chromosome aberrations.
- Alkylating agents of bases can change base-pairing properties.
- can also activate error-prone DNA repair processes.

Alkylating agents



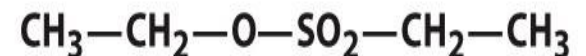
Di-(2-chloroethyl) sulfide

(Mustard gas)



Ethyl methane sulfonate

(EMS)



Ethyl ethane sulfonate

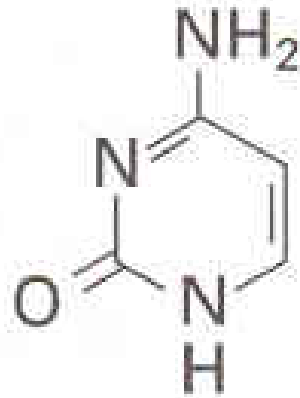
(EES)

Hydroxylamine

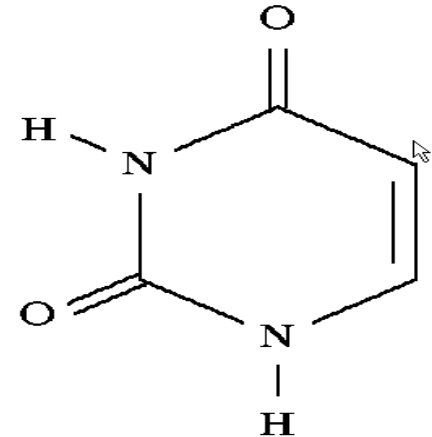
- Hydroxylamine is a hydroxylating agent.
- Hydroxylamine hydroxylates the amino group of cytosine and leads to $G:C \rightarrow A:T$ transitions.

5 Methyl Cytosine Deamination

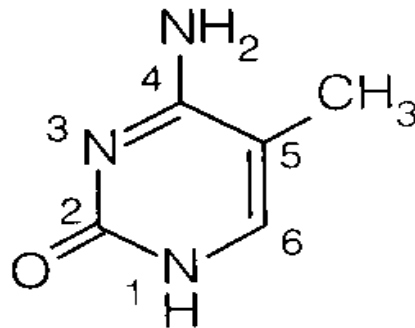
- Easily recognized and corrected
- What about 5-methyl cytosine?
- Is there a problem?
- Always remove T from a GT pair



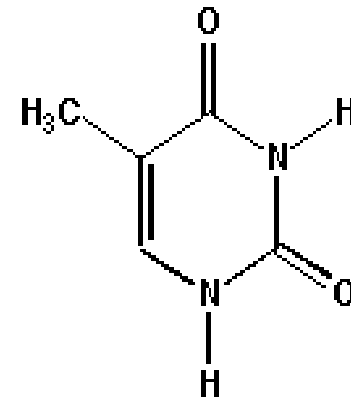
Cytosine



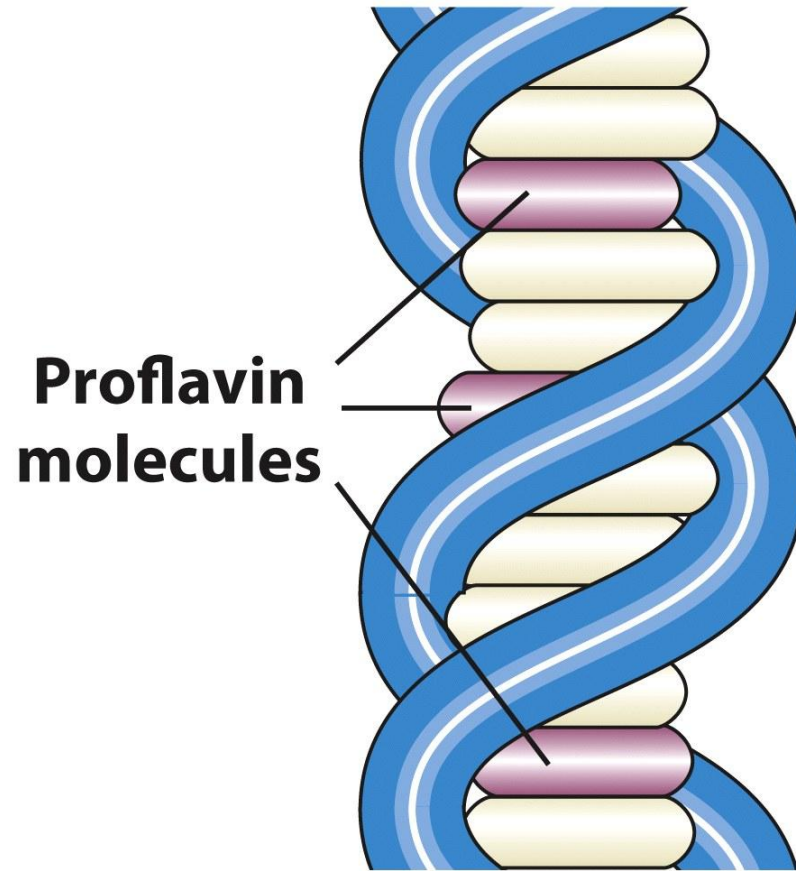
Uracil



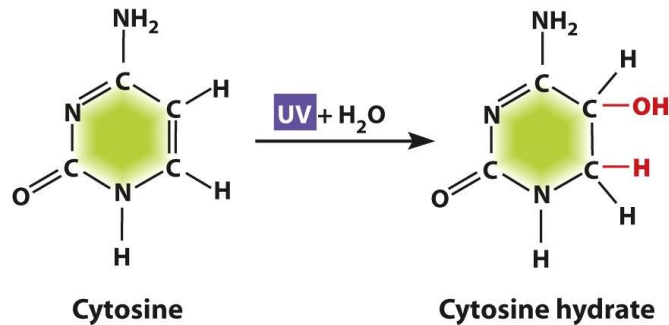
5-Methylcytosine



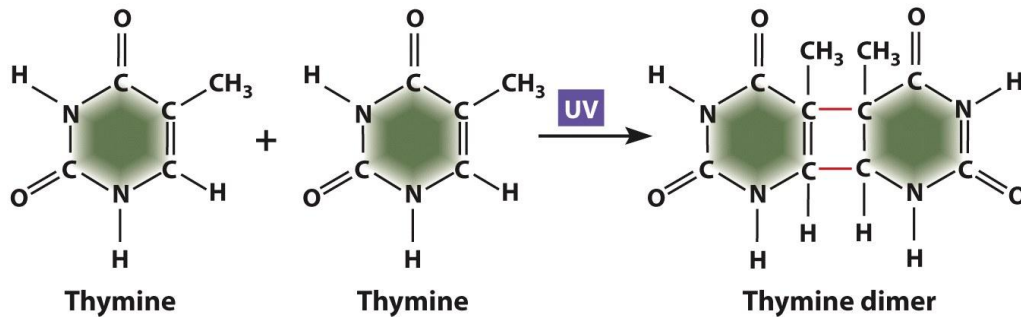
Intercalation of an Acridine Dye Causes Frameshift Mutations



Mutagenesis by Ultraviolet Irradiation



(a)



(b)

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- Hydrolysis of cytosine to a hydrate may cause mispairing during replication
- Cross-linking of adjacent thymine forms thymidine dimers, which block DNA replication and activate error-prone DNA repair mechanisms.

Mutation Generation passed on to daughter DNAs

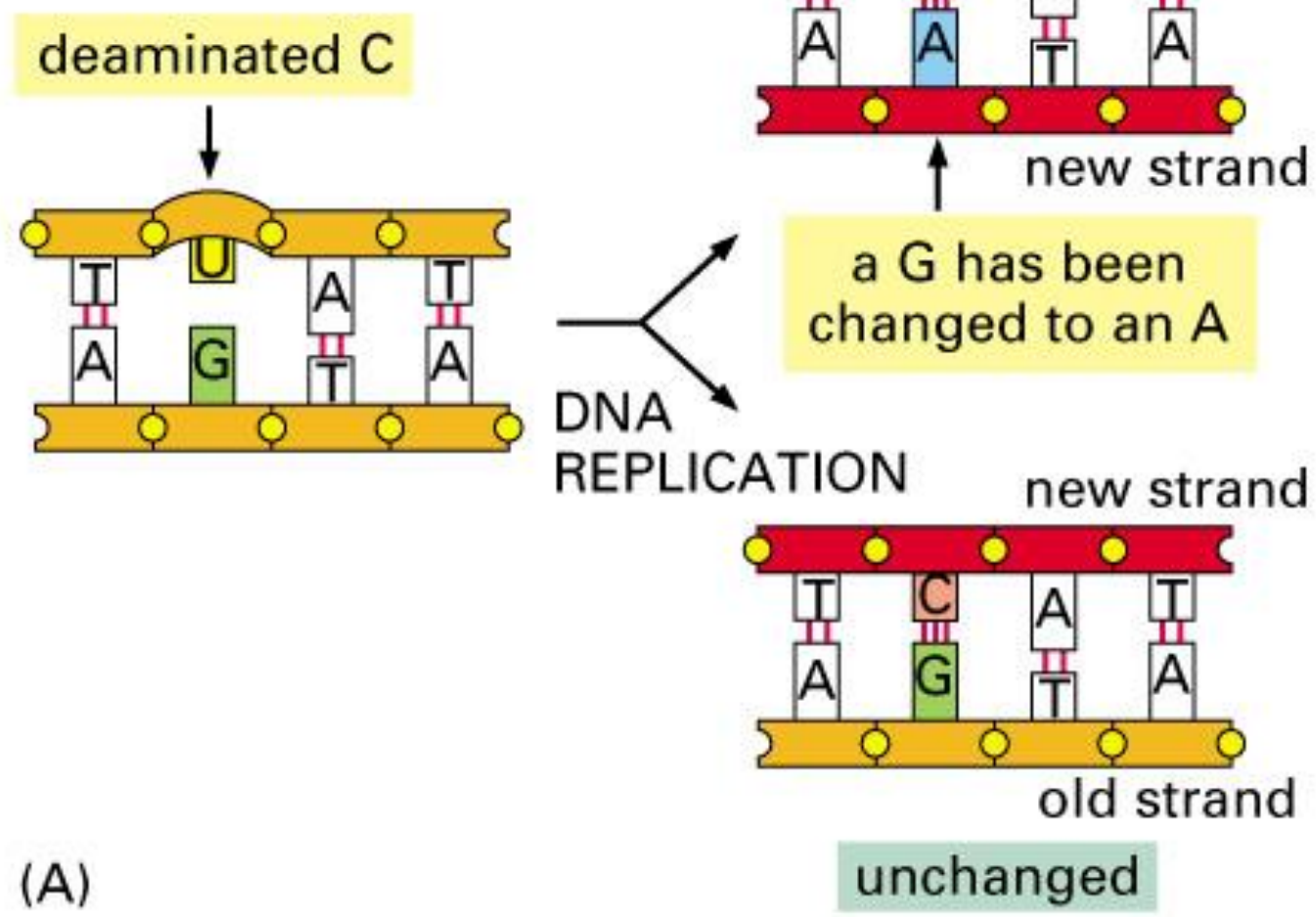
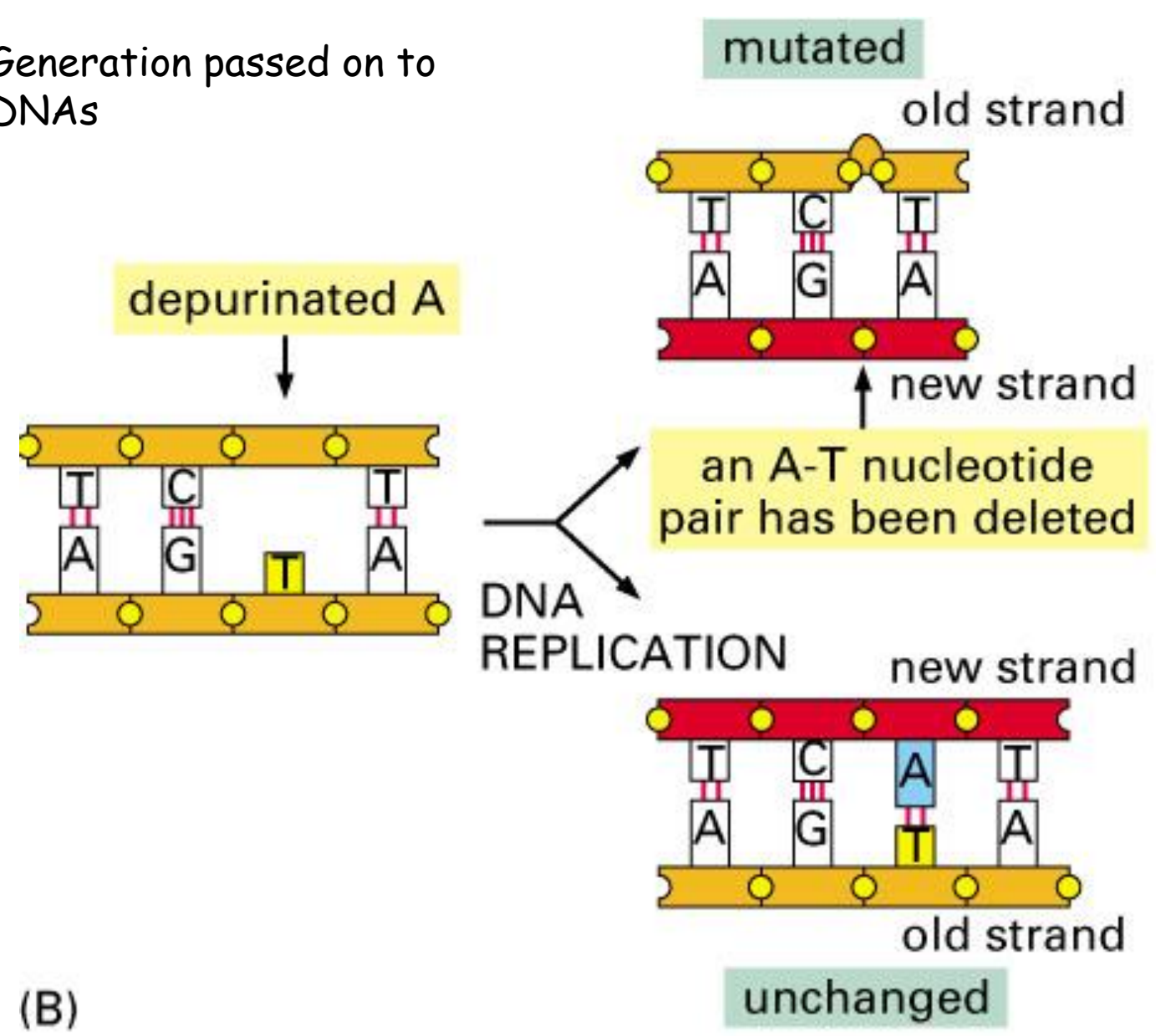


Figure 5-49 part 1 of 2. Molecular Biology of the Cell, 4th Edition.

Mutation Generation passed on to daughter DNAs



(B)

Figure 5-49 part 2 of 2. Molecular Biology of the Cell, 4th Edition.

Somatic Mutations

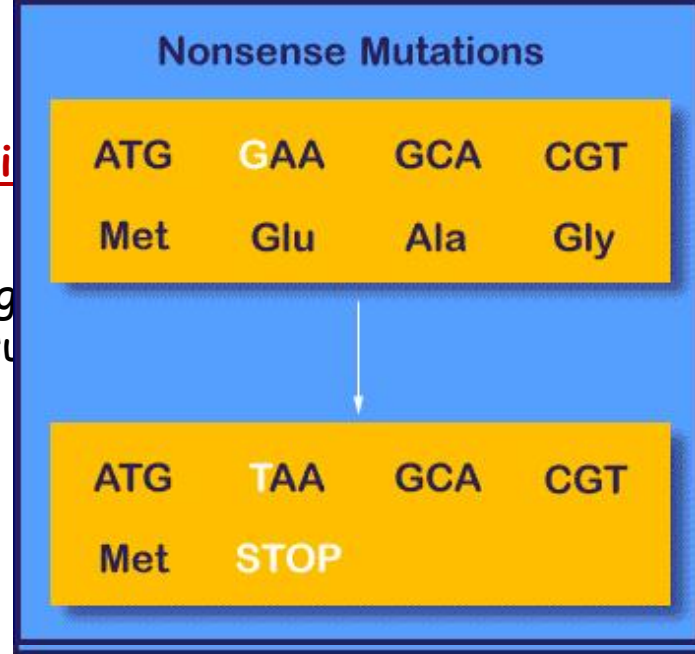
- Occur in cells of the body, excluding the germline
- Affects subsequent somatic cell descendants
- Not transmitted to offspring

Germline Mutations

- Mutations that occur in the germline cells
- Possibility of transmission to offspring

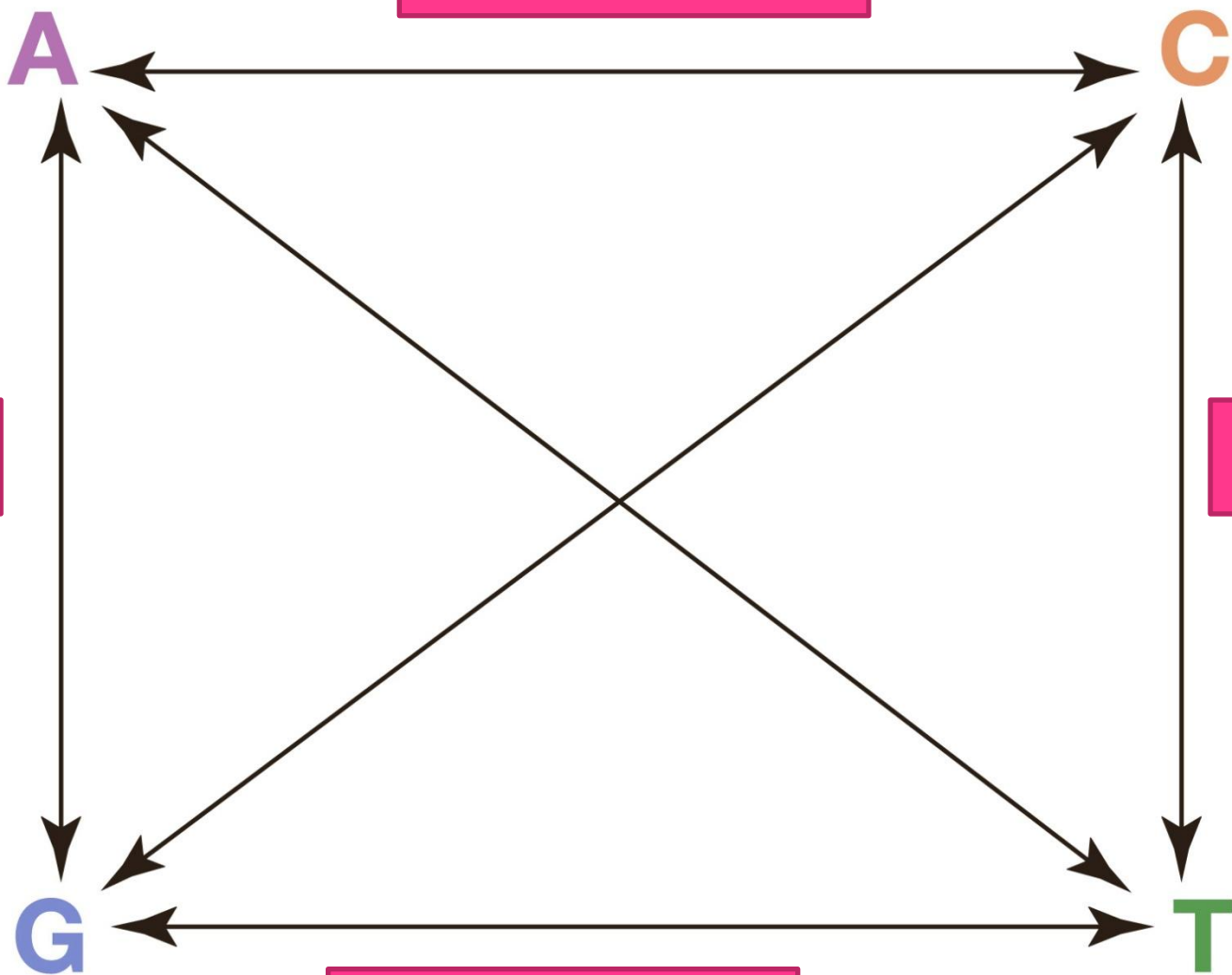
Gene mutation (alteri

- **Point mutation:** change of the following 3 resu



the mRNA can lead to any

- **i- Silent mutation:** i.e. the codon containing the changed base may code for the same amino acid. For example, in serine codon UCA, if A is changed to U giving the codon UCU, it still code for serine. See table.
- **ii- Missense mutation:** the codon containing the changed base may code for a different amino acid. For example, if the serine codon UCA is changed to be CCA (U is replaced by C), it will code for proline not serine leading to insertion of incorrect amino acid into polypeptide chain.
- **iii- Non sense mutation:** the codon containing the changed base may become a termination codon. For example, serine codon UCA becomes UAA if C is changed to A. UAA is a stop codon leading to termination of translation at that point.



DNA Repair I

(A) BASE EXCISION REPAIR

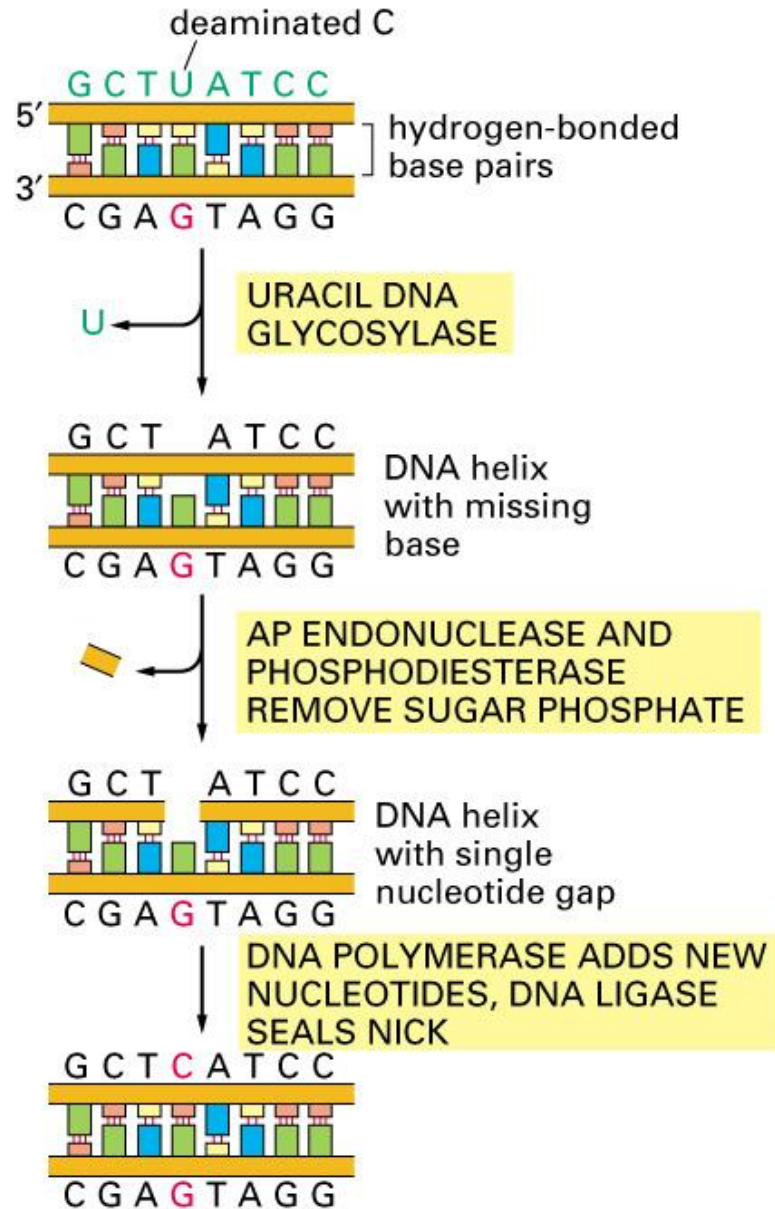


Figure 5-50 part 1 of 2. Molecular Biology of the Cell, 4th Edition.

DNA Repair II

(B) NUCLEOTIDE EXCISION REPAIR

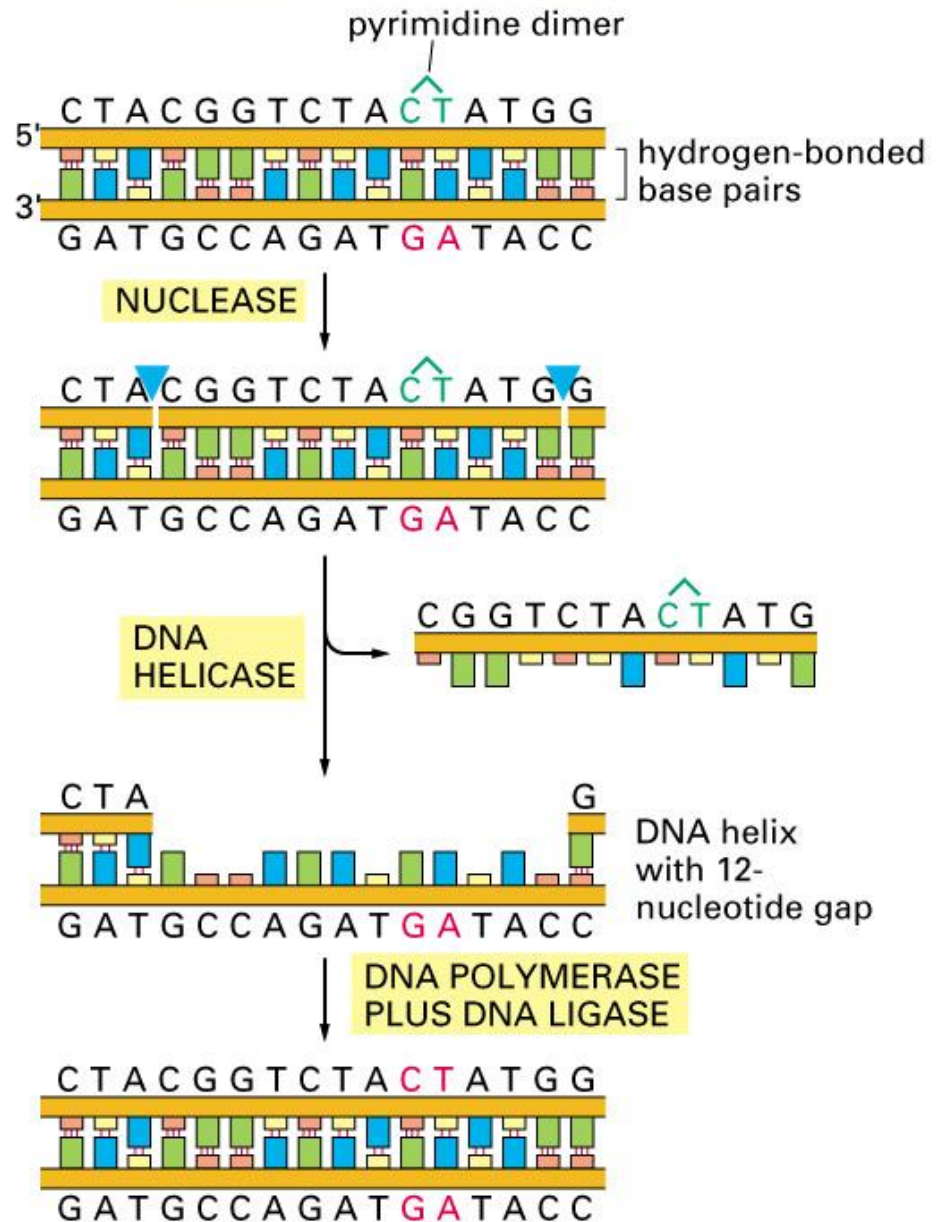


Figure 5-50 part 2 of 2. Molecular Biology of the Cell, 4th Edition.

Emergency DNA Repair for Double helix break

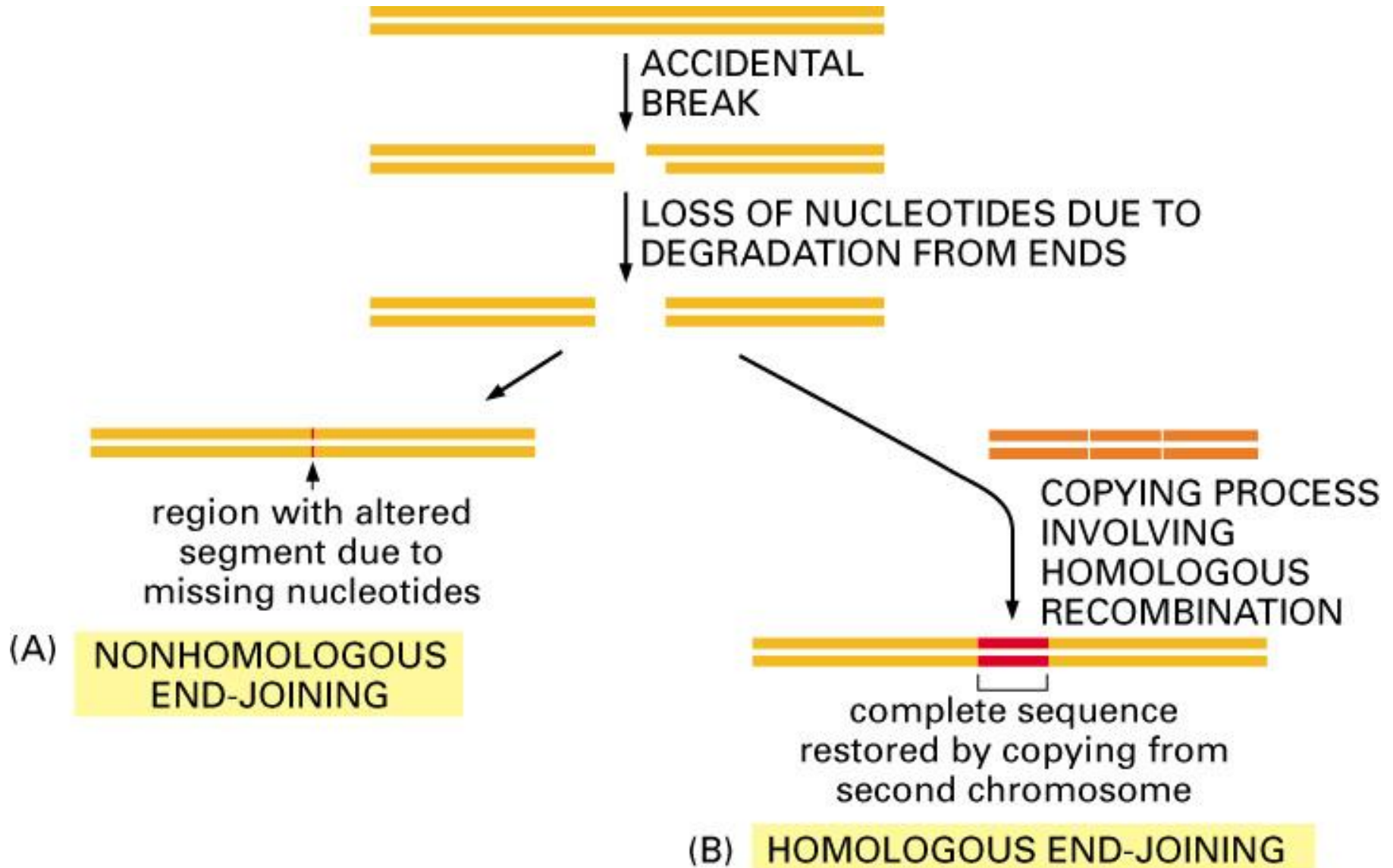


Figure 5-53. Molecular Biology of the Cell, 4th Edition.

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Proteins

Sequence Analysis

Taxonomy

Training & Tutorials

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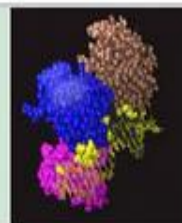
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Explore three-dimensional structures of proteins, DNA, and RNA molecules. Examine sequence-structure relationships, active sites, molecular interactions, biological activities of bound chemicals, and associated biosystems.



1 2 3 4 5 6 7 8

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Now Available: NCBI Insights Blog!
28 Jan 2013

NCBI has just released a new blog called *NCBI Insights*. Blog posts will provide an inside narrative to

Come to the NCBI Discovery Workshops on February 4&5!

16 Jan 2013

Spaces are still available for the free, 2 day Discovery Workshops to be held

IDH - Nucleotide - NCBI

www.ncbi.nlm.nih.gov/nuccore/?term=IDH

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NucleotideNucleotideIDHSearchSave searchLimitsAdvancedHelp

Display Settings:Summary, 20 per page, Sorted by Default orderSend to:Filter your results:

Found 5147 nucleotide sequences. Nucleotide (5089) EST (57) GSS (1)

Results: 1 to 20 of 5089<< First< PrevPage 1 of 255Next> Last>>

☐ Homo sapiens isocitrate dehydrogenase 1 (NADP+), soluble (IDH1), mRNA

1. 2,339 bp linear mRNA

Accession: NM_005896.2 GI: 28178824

GenBankFASTAGraphicsRelated Sequences

☐ Homo sapiens nucleophosmin (nucleolar phosphoprotein B23, numatrin) (NPM1), transcript variant 3, mRNA

2. 1,347 bp linear mRNA

Accession: NM_001037738.2 GI: 262331549

GenBankFASTAGraphicsRelated Sequences

☐ Homo sapiens nucleophosmin (nucleolar phosphoprotein B23, numatrin) (NPM1), transcript variant 2, mRNA

3. 1,362 bp linear mRNA

Accession: NM_199185.3 GI: 262331548

GenBankFASTAGraphicsRelated Sequences

☐ Homo sapiens nucleophosmin (nucleolar phosphoprotein B23, numatrin) (NPM1), transcript variant 1, mRNA

4. 1,449 bp linear mRNA

Accession: NM_002520.6 GI: 262331543

GenBankFASTAGraphicsRelated Sequences

☐ Azoarcus sp. KH32C DNA, complete genome

5. 5,081,166 bp circular DNA

All (5089)Bacteria (1277)INSDC (GenBank) (2970)mRNA (1734)RefSeq (2115)Manage Filters

Top Organisms [Tree]Homo sapiens (92)Giardia intestinalis (80)Pieris rapae (75)Mus musculus (61)Entamoeba histolytica (59)All other taxa (4723)More...

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Search detailsIDH[All Fields]

Nucleotide Nucleotide Search Limits Advanced Help

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Send:

Homo sapiens isocitrate dehydrogenase 1 (NADP+), soluble (IDH1), mRNA

NCBI Reference Sequence: NM_005896.2

FASTA Graphics

Go to:

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VERSION NM_005896.2 GI:28178824
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ORGANISM Homo sapiens
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AUTHORS Gerardo Valadez, J., Grover, V.K., Carter, M.D., Calcutt, M.W., Abiria, S.A., Lundberg, C.J., Williams, T.U. and Cooper, M.K.
TITLE Identification of Hedgehog pathway responsive glioblastomas by isocitrate dehydrogenase mutation
JOURNAL Cancer Lett. 328 (2), 297-306 (2013)
PubMed 23063752
REMARK GeneRIF: This study demonstrates that IDH mutation is a reliable surrogate marker for identifying malignant gliomas with an operant Hedgehog pathway.
REFERENCE 2 (bases 1 to 2339)
AUTHORS Jin, G., Reitman, Z.J., Duncan, C.G., Spasojevic, I., Gooden, D.M.,

Change region shown

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Articles about the IDH1 gene

Disruption of wild-type IDH1 suppresses D-2-hydroxyglutarate production [Cancer Res. 2013]

Hotspot mutations in H3F3A and IDH1 define distinct epigenetic and bic [Cancer Cell. 2012]

Prospective serial evaluation of 2-hydroxyglutarate, during treatment [Blood. 2012]

See all...

Pathways for the IDH1 gene

Abnormal conversion of 2-oxoglutarate to 2-

publications that are available for this gene. Please see the gene record to access additional publications.

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Transcript_exon_combination_evidence :: AF020038.1, AF113917.1
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COMPLETENESS: complete on the 3' end.

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Top of

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IDH (5089)

Nucleotide

See more...

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