

## UMUT GAZI, PhD

### SECTION 1 - PERSONAL DETAILS

<b>Title:</b> Asst. Prof. Dr.	<b>Forename:</b> Umut	<b>Surname:</b> Gazi
<b>Email address:</b> umut.gazi@hotmail.com, umut.gazi@neu.edu.tr		<b>Date of birth:</b> 12 April 1984

### SECTION 2 – ACADEMIC RELATED EXPERIENCE

#### **NEAR EAST UNIVERSITY**

**April 2012 – Present:** Lecturer at Medical Microbiology and Clinical Microbiology Department, Faculty of Medicine, Near East University (Cyprus).

### SECTION 3 – EDUCATION

#### **PhD in Immunology**

**September 2006 – December 2010:** UNIVERSITY OF NOTTINGHAM (UK).

**Research project:** Mannose receptor (MR) in macrophage biology. *Funded by the Medical Research Council (UK).*

**Project supervisor:** Dr. Luisa Martinez-Pomares.

#### **Project synopsis:**

- Investigated the possible effect of macrophage – collagen adhesion on MR expression and function as an endocytic receptor *in vitro*.
- Set up an *in vitro* model to study the mechanism responsible for the enhanced macrophage MR shedding in response to fungi.
- Identified the fungal cell wall component and the associated macrophage receptor as well as intracellular signaling involved in the fungi-mediated MR shedding.
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#### **Master of Research (MRes) in Biotechnology**

**September 2005 – September 2006:** UNIVERSITY OF KENT (UK).

**Modules studied include:** Molecular biology, analytical biotechnology, and statistics.

**Research project:** Notch signalling and nuclear positioning in *Drosophila* eye development.

**Project supervisor:** Dr. Marcus Allen.

#### **Project synopsis:**

- Showed that the Notch signalling pathway has a proofreading activity in nuclei localisation within the post-mitotic photoreceptors.
- Investigated the role of dynein-dynactin retrograde motor complex in maintaining correct nuclear positions in post-mitotic photoreceptors.

#### **BSc (Hons) in Molecular and Cellular Biology**

**September 2002 – July 2005:** UNIVERSITY OF KENT (UK). **68.6% (2.1) final grade.**

**Modules studies include:** Molecular and cellular biology, biochemistry, microbiology, organic chemistry, physiology, immunology, pharmacology, oncology, developmental biology, and neurology.

**Dissertation project:** Expression, purification and characterisation of *Mycobacterium Tuberculosis* proteins Rv0287 and Rv0288.

**Project supervisor:** Dr. Richard Williamson.

### **Foundation Course in Biological Sciences**

**September 2001 – June 2002: UNIVERSITY OF KENT (UK). 85% (1st) final grade.**

**Modules studied include:** Biology, chemistry, and mathematics

### **School**

**September 1998 – June 2001: TURK MAARIF KOLLEJI (CYPRUS). 88.2% final grade.**

## SECTION 4 – PUBLICATIONS AND MEETING ABSTRACTS

### **PUBLICATIONS**

- **Gazi U**, Rosas M, Singh S, Heinsbroek S, Haq I, Johnson S, Brown GD, Williams DL, Taylor PR, and Martinez-Pomares L. “*Fungal recognition enhances mannose receptor shedding through dectin-1 engagement*”. *Journal of Biological Chemistry* 2011; 286 (10): 7822-7829.

- **Gazi U**, and Martinez-Pomares L. “*Influence of the mannose receptor in host immune response*”. *Immunobiology* 2009; 214 (7): 554-61.

### **MEETING ABSTRACTS**

-**Gazi U**, Heinsbroek S, Taylor PR, Williams DL, Brown GD, and Martinez-Pomares L. “*Dectin-1-mediated signaling enhances mannose receptor shedding*”. Poster presentation at the 2<sup>nd</sup> Congress of European Immunology in Berlin, Germany, 2009.

-**Gazi U**, Williams DL, Brown GD, and Martinez-Pomares L. “*Dectin-1-mediated signaling enhances mannose receptor shedding*”. Poster presentation at the British Society of Immunology (BSI) Congress in Glasgow, UK, 2008.

## SECTION 5 – RESEARCH TECHNIQUES, PERSONAL SKILLS AND COMPETENCES

### **RESEARCH TECHNIQUES**

**Expertise:** RT-PCR; cell RNA and DNA extraction, isolation and quantification; cDNA synthesis; ELISA (sandwich, competitive, and indirect); cell culture (mammalian and immortalized cell lines); immunocytochemistry; light and fluorescence microscopy; western blotting of cell lysates and culture supernatants; preparation of cell lysates and protein quantification; inhibition assays; flow cytometry to screen the level of ligand internalization and antigen expression; intraperitoneal injection of mice; isolation of bone marrow and thioglycollate elicited macrophages from mouse; preparation of extracellular matrix protein-coated plastic plates.

**Familiar:** Gelatin zymography; *in situ* hybridization; protein purification by column chromatography; dissection of *Drosophila* larvae organs; *Drosophila* culture; fluorescence spectroscopy; circular dichroism spectroscopy; mass spectroscopy; restrictive enzyme digestion; gene cloning and bacterial transformation.

## **PERSONAL SKILLS AND COMPETENCES**

**Languages:** English and Turkish (Fluent), currently attending Greek language classes.

**Computer Skills:** PowerPoint, Excel, Word, WinMDI, Weasel, Statistical analysis of biological data, Adobe Photoshop.

## **SECTION 6 – OTHER RELATED EXPERIENCE**

<b>Employer</b>	<b>Position Held</b>	<b>Key Responsibilities</b>	<b>Dates - From/To</b>
University of Kent Biosciences Department	Teaching Assistant	Demonstrating in laboratory practical sessions	09/2005- 09/2006 (Part-time)
Nalbantoglu State Hospital (Nicosia / Cyprus)	Volunteer	Blood and urine testing, syphilis, AIDS, Hepatitis and Sickle Cell Anaemia testing and the CBC test	07/2005- 08/2005
Nalbantoglu State Hospital (Nicosia / Cyprus)	Volunteer	Blood and urine testing, syphilis, AIDS, Hepatitis and Sickle Cell Anaemia testing and the CBC test	07/2004- 08/2004

## **SECTION 7 – AWARDS, GRANTS AND INTERESTS**

**-Travel Grant**, British Society of Immunology to attend the 2<sup>nd</sup> Congress of European Immunology in Berlin, Germany (2009).

**-Best Poster Presentation Prize**, MOL Postgraduate Research Day at the University of Nottingham, UK (2008).

**-MRC studentship** for PhD research (2006-2009).

**-Scholarship/Bursary**, The University of Kent, UK (2002).